



Physiological and ecological tipping points caused by ocean acidification

Christopher E. Cornwall¹, Steeve Comeau², Ben P. Harvey³

¹ School of Biological Sciences and Coastal People Southern Skies Centre of Research Excellence, Victoria University of Wellington, Wellington, 6012, New Zealand

² Sorbonne Université, CNRS-INSU, Laboratoire d'Océanographie de Villefranche, F-06230 Villefranche-sur-Mer, France

³ Shimoda Marine Research Center, University of Tsukuba, Shimoda, Shizuoka, Japan

Correspondence to: Christopher E. Cornwall (Christopher.cornwall@vuw.ac.nz)

10 Abstract.

Ocean acidification is predicted to cause profound shifts in many marine ecosystems by impairing the ability of calcareous taxa to calcify and grow, and by influencing the photo-physiology of many others. In both calcifying and non-calcifying taxa, ocean acidification could further impair the ability of marine life to regulate internal pH, and thus metabolic function and/or behaviour. Identifying tipping points at which these effects will occur for different taxa due to the direct impacts of ocean acidification on organism physiology is difficult and they have not adequately been determined for most taxa, nor for ecosystems at higher levels. This is due to the presence of both resistant and sensitive species within most taxa. However, calcifying taxa such as coralline algae, corals, molluscs, and sea urchins appear to be most sensitive to ocean acidification. Conversely, non-calcareous seaweeds, seagrasses, diatoms, cephalopods, and fish tend to be more resistant, or even benefit from the direct effects of ocean acidification. While physiological tipping points of the effects of ocean acidification either do not exist or are not well defined, their direct effects on organism physiology will have flow on indirect effects. These indirect effects will cause ecologically tipping points in the future through changes in competition, herbivory and predation. Evidence for indirect effects and ecological change is mostly taken from benthic ecosystems in warm temperate–tropical locations *in situ* that have elevated CO₂. Species abundances at these locations indicate a shift away from calcifying taxa and towards non-calcareous at high CO₂ concentrations. For example, lower abundance of corals and coralline algae, and higher covers of non-calcareous macroalgae, often turfing species, at elevated CO₂. However, there are some locations where only minor changes, or no detectable change occurs. Where ecological tipping points do occur, it is usually at locations with naturally elevated pCO₂ concentrations of 500 μatm or more, which also corresponds to just under that concentrations where the direct physiological impacts of ocean acidification are detectable on the most sensitive taxa in laboratory research (coralline algae and corals). Collectively, the available data support the concern that ocean acidification will most likely cause ecological change in the near future in most benthic marine ecosystems, with tipping points in some ecosystems at as low as 500 μatm pCO₂. However, much more further research is required to more adequately quantify and model the extent of these impacts in order to accurately project future marine ecosystem tipping points under ocean acidification.



1 Introduction

Ocean acidification is the process of increasing absorption of atmospheric CO₂ by the surface seawaters, leading to a decrease in pH and shift in the speciation of dissolved inorganic carbon (DIC). As a result, future seawater concentrations of HCO₃⁻, H⁺, and CO₂ will be higher, while CO₃²⁻ will be lower. These changes in seawater carbonate chemistry will have complex biological consequences, as all four parameters mentioned previously hold physiological significance for various marine taxa (Hurd et al., 2019). The direct effects of ocean acidification manifest mostly through three mechanisms: 1) negatively impacting calcification (and recruitment of calcareous organisms); 2) altering photo-physiology; and 3) affecting acid-base regulation (i.e., internal pH regulation) or the energy expended in this process. These effects are usually subtle and have relatively minor direct negative effects when compared to stressors like marine heatwaves. However, the cumulative impacts of these physiological changes on individual species will result in more significant ecological effects. Comprehensive meta-analyses on the impacts of ocean acidification on biological processes and ecological outcomes support these statements (Leung et al., 2022; Kroeker et al., 2013b). However, these analyses rarely assess changes in processes at specific time points or at *p*CO₂ values corresponding to such time points. Instead, they usually determine whether standardised responses are different from zero. Thus, in this study, we evaluate when key responses in different taxa would differ from zero and provide *p*CO₂ values of when this typically occurs, if it is possible to make such statements. We also provide background information describing the key physiological and ecological impacts of ocean acidification.

2 Direct impacts on key physiological processes

2.1 Calcification

Calcium carbonate minerals precipitate or dissolve relative to their saturation state (Ω), which is determined by [Ca²⁺] multiplied by [CO₃²⁻] and divided by *K*_{sp}. *K*_{sp}, being dependent on seawater temperature, salinity, and pressure (Morse and Mackenzie, 1990). Consequently, under ocean acidification, declines in [CO₃²⁻] and Ω lead to reduced mineral precipitation in seawater. In theory, minerals like calcite have higher Ω values than aragonite, while aragonite has a higher Ω than high Mg calcite (Andersson et al., 2008; Bathurst, 1972; Feely et al., 2004). Initial experimental work demonstrated that simulated ocean acidification reduces calcification in most taxa (Ries et al., 2009). However, there are also instances of resistant species or entire taxa (Kroeker et al., 2013b; Leung et al., 2022). Thus, the initial assumption was largely that ocean acidification reduces calcification by decreasing Ω . Indeed, taxa that precipitated lower Ω minerals tended to be more strongly impacted by ocean acidification (Kroeker et al., 2013b). CO₃²⁻ was more highly correlated than other components of seawater carbonate chemistry in experiments where the independent role of pH, HCO₃⁻ and/or CO₃²⁻ were tested, and teased apart independently (Comeau et al., 2013a; Schneider and Erez, 2006). However, calcification generally occurs internally (Ries, 2011). In organisms such as corals, it takes place several tissue layers deep (Allemand et al., 2004), while in coralline algae, it occurs



65 within the cell wall, one to several cells deep (Cornwall et al., 2017a). This internal location is referred to as the calcifying fluid (Mcculloch et al., 2012a; Mcculloch et al., 2012b). Notably, there are no known biological transporters of CO_3^{2-} , and it remains membrane impermeable in all organisms. Therefore, it is more likely that increases in external $[\text{H}^+]$ drive the observed declines in calcification rates observed in corals, a process offset by the provision of greater DIC under ocean acidification (Jokiel, 2013).

To calcify, marine organisms must create conditions that favour the precipitation of calcium carbonate at the site of calcification. Most calcifying organisms actively adjust their internal chemistry to initiate the inorganic precipitation of calcium carbonate by increasing the pH and ensuring a consistent supply of essential ions required for calcification (CO_3^{2-} , HCO_3^- , Ca^{2+} , Mg^{2+}) (Mcculloch et al., 2017; Decarlo et al., 2018). This is done by actively removing protons from the site of calcification via specific cellular transporter. The inorganic carbon utilized for calcification originates from a combination of metabolic CO_2 , HCO_3^- transported to the site of calcification via cellular transporters, and ions transported via paracellular pathways (e.g., Venn et al., 2020). Additionally, marine organisms have developed specific proteins that aid in maintaining elevated calcium carbonate precipitation (Marin et al., 2007; Drake et al., 2013). While the general principles of calcification are similar across many taxa, the response to ocean acidification is highly taxa and species-specific. Here we will focus on three of the most studied calcifying taxa: corals, molluscs, and calcifying macroalgae. These three taxa are also identified as the most at risk from the effects of ocean acidification.

Corals have received significant attention in studies related to ocean acidification. However, results regarding a potential tipping point in their response to ocean acidification are inconclusive, and the shape of the relationship between calcification and pH (or pCO_2) is still a subject of debate. Early research suggested a linear response of corals growth to pH (Doney et al., 2009; Anthony et al., 2008), but some studies challenged this finding, showing non-linear relationships between calcification and pH (or Ω) (e.g., Ries et al., 2009). It is however important to note that most of these studies had limited treatment levels, making it difficult to assess the shape of the ocean acidification to calcification relationship and detect tipping points. More recent studies, using a wider range of treatments (Comeau et al., 2013b, 2014) and species (Okazaki et al., 2017), experimentally demonstrated that the response of coral calcification to pCO_2 is generally linear. Linear relationships have been considered the best representation of the coral response of corals to ocean acidification in recent meta-analyses (Cornwall et al., 2021; Kornder et al., 2018). Therefore, there is no clear tipping point in the response of corals to ocean acidification, as their response is generally linear, highly species-specific, and influenced by other factors such as light, temperature, and feeding. At the reef level, studies indicate a linear decline in calcification with the saturation state, but the magnitude of this decrease (i.e., the slope) varies among reefs and likely depends on community composition. Some studies project a shift to net



dissolution when Ω decreases below 2, a level that could be reached by the end of the century (Andersson and Gledhill, 2013; Albright et al., 2018).

Molluscs, especially commercial ones, have also been extensively studied in relation to their response to ocean acidification due to their economic importance. Previous reviews on the effects of ocean acidification on molluscs, though conducted several years ago, revealed that the response of adults and juveniles varies significantly among species and even within species (Gazeau et al., 2013; Parker et al., 2013). Similar to corals, the response of molluscs to ocean acidification is strongly influenced by their environmental history and interactions with other environmental factors (Falkenberg et al., 2019; Thomsen and Melzner, 2010). Different mollusc species precipitate various forms of calcium carbonate, such as calcite, aragonite, or high Mg-Calcite, which are known to respond differently to ocean acidification. Moreover, there is evidence suggesting that molluscs could undergo transgenerational acclimation to ocean acidification, potentially mitigating the negative effects of decreasing pH (Parker et al., 2021). As a result, determining a tipping point at which mollusc calcification and growth might collapse proves to be extremely challenging. Instead, it is more likely that some species will experience a steady decrease in calcification and shell strength, while others may thrive in lower pH conditions. The complexity of these responses makes it difficult to predict the overall impact of OA on molluscs with certainty.

Coralline algae are extremely sensitive to the effects of ocean acidification, with their calcification rates, cover in the field at natural CO₂ sites, recruitment in laboratory and in situ work, and internal pH at the site of calcification all negatively impacted by ocean acidification (Cornwall et al., 2022). Calcification rates of coralline algae are negatively affected 75% of the time, with in most instances a linear decrease of calcification with decreasing pH. However, like in molluscs, greater resistance to ocean acidification has been demonstrated in coralline algae grown for multiple generations (6 generations) under conditions simulating the effects of ocean acidification, versus those from controls (Cornwall et al., 2020; Moore et al., 2021). Additionally, some species display natural tolerance to ocean acidification (Cornwall et al., 2018; Cornwall et al., 2017a). Although the physiological mechanisms via which this tolerance is achieved is relatively unexplored, there are possible links between greater upregulation of pH within the site of calcification and maintenance of calcification under ocean acidification. Irrespective of these tolerance species, it is likely coralline algae will be among the first taxa to clearly show negative impacts of ocean acidification, as on average they are universally sensitive. This will cause large flow on effects at ecological levels (see section below on this), as they play important roles in maintaining coral reef growth and cementation (Cornwall et al., 2023), and act as crucial settlement substrates for invertebrate larvae such as corals, abalone and sea urchins (Roberts, 2001; Fabricius et al., 2017).

Declines in calcification within most taxa due to ocean acidification is expected, but determining a specific tipping point for when this decline will occur is a complex task. The complexity arises from the diverse approaches used in past research, the



different atmospheric CO₂ concentrations projected under various climate change scenarios, and the lack of alignment between meta-analyses and these atmospheric CO₂ concentration scenarios. We summarise the past attempts at meta-analyses, or pseudo-meta-analyses in Table 1. Notably, there is a significant overlap between the majority of past meta-analyses (Hendriks et al., 2010; Kroeker et al., 2013b; Kroeker et al., 2010; Harvey et al., 2013; Kornder et al., 2018) and more recent meta-analyses (Leung et al., 2022; Cornwall et al., 2021; Cornwall et al., 2022). Leung et al. offer a more detailed description of the responses of various calcifying taxa to ocean acidification compared to most prior meta-analyses. However, they group all relevant studies necessary to identify tipping points into a single category (~pH 7.9 to pH 7.6). Additionally, they group physiologically different calcifying algae that are not grouped in some other recent analyses on coralline algae [and corals], which attempt to determine temporal differences in effect sizes (Cornwall et al., 2021; Cornwall et al., 2022). Furthermore, Wittmann and Pörtner (2013), although not a true meta-analysis, provide detailed and interesting breakdowns of the impacts of different pH range values on calcification rates of various animal taxa. To estimate calcification and growth rates of most taxa, we consider them largely interchangeable since the ability to create new skeletal material is essential for inorganic growth and size increases. Therefore, for our estimates, we draw on all recent relevant research, including the work of Wittmann and Pörtner.

Evidence strongly suggests that the calcification rates of coccolithophores, pteropods, coralline algae, *Halimeda* spp., corals, bivalves, and gastropods are likely to experience significant decreases due to ocean acidification by the end of the century under RCP8.5 emissions scenarios or similar conditions (e.g., ~pH 7.63 and 936 μatm CO₂) (Leung et al., 2022; Cornwall et al., 2022). However, the evidence remains unclear for foraminifera, calcifying sponges, bryozoans, crustaceans, polychaetes, and echinoderms, as there is mixed evidence presently available (Leung et al., 2022). Specifically, coralline algal calcification rates significantly decline from present-day rates within ocean acidification scenarios that simulate RCP4.5 2050 and above (e.g., 538 μatm), whereas corals experience similar departures around 2050 RCP8.5 (e.g., 572 μatm) (Cornwall et al., 2021). Though not equivalent, the point at which more than 50% of studies find negative impacts on all measures of organism fitness is around 780 to 840 μatm for molluscs and echinoderms respectively, and above 2000 μatm for crustaceans (Wittmann and Pörtner, 2013). Although not all of these measurements directly involve calcification, this highlights the difficulty in determining physiological tipping points for changes in calcification rates for specific taxa, given the available and disparate research.

2.2. Photo-physiology

Most macroalgae and all phytoplankton species have the ability to directly take up HCO₃⁻ from seawater during photosynthesis, converting it into complex sugars (Raven et al., 2011; Raven et al., 2002a). Similarly, all marine phototrophs can passively take up CO₂ through diffusion for the same process (Raven et al., 2012), with some of them relying solely on this diffusive



uptake. In corals, symbiotic dinoflagellates use DIC within the coral tissues through the same process (Raven et al., 2020). The active uptake of HCO_3^- using pumps, symports, antiports is referred to as a CO_2 concentrating mechanism (CCM) (Raven et al., 2011). The evolution of CCM was believed to be a response to the low concentrations and slow diffusivity of CO_2 in seawater compared to air (Raven et al., 2002b; Raven et al., 2008), where ~90% of seawater DIC is in the form of HCO_3^- and only about 1% is in the form of CO_2 . Therefore, using HCO_3^- was considered to theoretically overcome DIC limitation. However, this reasoning is simplistic. This is noted in the various works by Raven and colleagues (e.g., Raven et al., 2005; Raven and Beardall, 2014, 2016), and we summarise here. There are various types of CCMs that differ in their efficiency in natural seawater. Although seawater pH typically is around ~8.05, it can be higher in regions with high photosynthetic uptake of DIC and high water retention (Rivest et al., 2017). In these habitats, CO_2 concentrations would be extremely small and species utilising diffusive CO_2 uptake would be at a competitive disadvantage over those with CCMs (Hepburn et al., 2011). In additions to true CCMs, there are various associated external and internal carbonic anhydrase enzymes that collectively enhance the diffusive CO_2 uptake externally, or convert internal CO_2 into HCO_3^- , thereby regulating internal CO_2 levels and maintaining internal pH. The efficiency and production of these enzymes can vary between species. Moreover, both the creation of CCMs and carbonic anhydrase enzymes require energy and nutrients. As a result, possessing a CCM does not guarantee that a species is not DIC limited in the habitats it currently occupies. Increasing CO_2 (and possibly HCO_3^-) could potentially alleviate DIC limitation in marine phototrophs that are constrained by DIC availability. However, quantifying the exact benefits that these organisms would receive is challenging because there has been no comprehensive effort to study this by subjecting organisms with well-known CCMs and carbonic anhydrases to seawater conditions resembling ocean acidification. Nevertheless, it has been observed that several DIC-limited species do show an increase in their photosynthetic rates and/or growth rates in response to elevated DIC or ocean acidification (Kroeker et al., 2010; Harvey et al., 2013).

Increasing CO_2 concentrations could also lead to species with a CCM using additional CO_2 when the external CO_2 concentration is higher, thus downregulating their CCM (Hepburn et al., 2011). The potential benefits of this downregulation might be minimal, but it does appear as though both marine and freshwater macroalgae at elevated CO_2 concentrations either downregulate their CCM or switch the ratio of HCO_3^- to CO_2 uptake (Cornwall et al., 2017b; Maberly et al., 2014). Another possible outcome is that higher external $[\text{CO}_2]$ levels could reduce the leakage of internal CO_2 back into seawater, saving energy and possibly nutrients. However, the specific impacts of these processes under ocean acidification are challenging to predict and likely depend on local irradiance availability and nutrient concentrations relative to the specific species' requirements.

Photosynthetic efficiency (F_v/F_m) is observed to decline in many taxa (e.g., corals, seaweeds) under ocean acidification (Leung et al., 2022; Cornwall et al., 2022), but the significance of these decreases as stress indicators is debatable. For instance, declines in F_v/F_m of approximately 0.4 or more are typically associated with acute heat stress (e.g., Schoepf et al., 2019),



185 whereas declines caused by simulated ocean acidification are usually on the order of 0.02. Therefore, it is possible that these declines are in response to non-stress related increases in cellular CO₂ or decreased pH, affecting the functioning of the photosystem II. Further research is needed to understand the physiological and molecular mechanisms that underpin these responses in various phototrophic organisms and to determine if they indeed represent significant changes that would impact future photosynthetic species in a future high CO₂ ocean.

190 Meta-analyses highlight that ocean acidification will impact the photo-physiology of marine species. However, due to the complexity of the underlying processes (as highlighted above), there is considerable variability within and between taxa, and limited data on photosynthetic responses. Consequently, it becomes even more challenging to pinpoint a tipping point at which changes in photosynthetic rates would occur. On average, ocean acidification seems to have a positive impact on coccolithophore photosynthetic rates, but there is no average detectable effect in foraminifera, symbiont bearing sponges, corals, and calcifying algae (Leung et al., 2022). However, both corals and coralline algae show declines in photosynthetic efficiency, and there is evidence of both strong positive and negative impacts on their photosynthetic rates (Leung et al., 2022; 195 Cornwall et al., 2022). The underlying physiological mechanisms for these negative observations require further investigation. Older meta-analyses also indicate positive effects of ocean acidification on seagrass, diatom, and non-calcareous macroalgal growth and photosynthesis, though with varied responses (see Table 1) (Harvey et al., 2013; Kroeker et al., 2013b).

2.3 Internal pH regulation

200 Ocean acidification also presents significant changes for the regulation of internal pH in different tissues or fluids of many calcifying taxa. This is evident in corals (Mcculloch et al., 2012b; Venn et al., 2013), coralline algae (Cornwall et al., 2017a), *Halimeda* spp. (Comeau et al., 2019), echinoderms (Stumpp et al., 2012), and crustaceans (Carter et al., 2013) for example. However, some of these species show an increased ability to upregulate internal pH at sites of calcification, allowing them to maintain calcification under simulated ocean acidification conditions. As a result, relying solely on meta-analyses to predict when specific species will be impacted by ocean acidification may have limitations. That being said, meta-analyses do indicate 205 significant impacts of OA on pH regulation in certain echinoderms and corals (Leung et al., 2022), as well as in coralline algae (Leung et al., 2022; Cornwall et al., 2022). However, the extent to which OA affects pH within different components of various taxa remains unexplored, and this could potentially explain some of the variability in responses even among non-calcifying taxa, such as many seaweeds. This is a vital controller of organism metabolism and fitness, but further research on this topic is required before tipping points could be identified.



210 3. Changes at naturally high CO₂ locations

Research on the potential effects of ocean acidification on marine organisms highlight the connection between increasing atmospheric and oceanic CO₂ levels, changes in carbonate chemistry, and their impacts on marine organisms. As highlighted previously, numerous experimental studies have documented significant impacts of future ocean acidification on diverse aspects of individual species' physiology, life history, and ecology, as well as on populations. However, when attempting to
215 assess the impact of ocean acidification at higher levels of biological organisation, such as community structure, food web dynamics, and ecosystem function, uncertainty arises. Particular difficulties arise when attempting to predict the outcomes of interactions among numerous species. Nevertheless, all these individual responses contribute to ecosystem change, and so ocean acidification is anticipated to lead towards changes in the structure, composition, and functioning of marine ecosystems.

220 Ecosystem changes, although usually gradual, can reach tipping points where they undergo sudden and significant shifts, leading to alterations in the structure and function of biological communities. These regime shifts are of particular concern due to the fact that the newly established habitats are primarily composed of species that have lower ecological, functional, and human value when compared to the habitats they have replaced. Examples of this include the extensive displacement of key foundation species (e.g., coral reefs and kelp forests) by simplified degraded ones like turf algae-dominated systems (e.g.,
225 Knowlton, 1992; Moy and Christie, 2012; Wernberg et al., 2016). Ecosystem changes associated with climate change are evident today in various ecosystems and biogeographic regions, often associated with temperature. However, these changes are often complex, involving concurrent shifts in multiple environmental factors and processes. On the other hand, the direct attribution of ongoing ecosystem changes, or specific elements, to anthropogenic ocean acidification is far more difficult to establish.

230 Seasonal upwelling regions, such as the California coast, offer valuable insights into the contemporary impacts of altered carbonate chemistry. In these areas, nutrient-rich waters from deeper layers of the ocean, which are also characterised by high CO₂ and low pH, are transported to the surface. This process provides examples of how low pH waters can bring about ecosystem changes by impacting early life stages of economically and ecologically important species (e.g., Pacific Shellfish
235 species) (Feely et al., 2008). Another contemporary example includes coral reefs in the Equatorial Upwelling System, such as those found in the Galápagos and Cocos Islands, which exhibit low species diversity and limited development of carbonate reef frameworks attributed to the low pH/aragonite saturation levels in the upwelling waters of the region (Glynn, 2001; Manzello, 2010; Manzello et al., 2008). One of the issues in the attribution of ecosystem change to ocean acidification (beyond the confounding effects of other environmental factors) is associated with the difficulty in the large-scale monitoring of
240 carbonate chemistry. However, a study examining artificial ocean alkalisation on a localised scale yielded important findings (Albright et al., 2016). It showed that increasing alkalinity to pre-industrial levels resulted in an increase in net community calcification at a coral reef on One Tree Island, Great Barrier Reef. This study therefore suggests that contemporary



communities are experiencing a decline in net community calcification compared to pre-industrial conditions, indicating that coral reef growth may potentially already be impaired by ocean acidification. Conversely, the addition of CO₂ to a coral reef at the same location caused declines in net community calcification rates beyond those expected to occur due to the direct impacts on coral (Albright et al., 2018). This could be due to resident coralline algae or carbonate sediments, that are high in abundance in some areas on this reef and more prone to dissolution at rates much higher than their calcification rates. While contemporary examples, such as regions of seasonal upwelling, promote our understanding of the current impacts of altered carbonate chemistry, understanding the potential future impacts of ocean acidification remains crucial.

Research efforts are also currently underway into the long-term effects of ocean acidification on ecosystem-level processes using natural analogues (Figure 1). Natural analogues for ocean acidification predominantly involve a natural, localised change in carbonate chemistry conditions that mimic future ocean acidification conditions which can be compared with an adjacent region under contemporary conditions. These include CO₂ seeps, associated with volcanic CO₂ emissions, and semi-enclosed bays (e.g., semi-enclosed lagoons, and mangrove estuaries), where certain ecologically significant parameters for marine life (carbonate chemistry, temperature, oxygen) closely resemble or even surpass the projected conditions anticipated for the end-of-the-century. Since marine communities and ecosystems are home to a wide range of species, that each interact with each other, these research approaches importantly enable researchers to simultaneously examine the response of a naturally-assembled community as a whole; regardless of whether they are investigating a specific population, functional group or the community as a whole. While not exact replicas of future conditions, these natural laboratories present unique opportunities to investigate the mechanisms by which ecosystems may adapt and respond to the challenges of climate change.

In early studies utilising CO₂ seeps (Hall-Spencer et al., 2008), the loss of vulnerable calcifying organisms and the potential benefit observed in certain non-calcified primary producers (such as seagrass and algae) were noted. However, it was also observed that some calcifying organisms managed to survive while certain non-calcifying primary producers were being lost. This significant finding emphasised the limitations of solely considering the direct physiological impacts of ocean acidification when predicting community-level responses, highlighting the need for a comprehensive understanding of the ecological interactions within a community (Fabricius et al papers). Ongoing research at a number of different CO₂ seeps has consistently documented shifts in habitat structure and community composition along the natural *p*CO₂ gradients. These shifts often favored simplified systems with reduced biodiversity and less ecological complexity over larger habitat-forming species (Sunday et al., 2016). Examples include transitions from canopy-forming brown algae to turf (Ischia, Italy), hard corals to soft corals (Iwotorishima, Japan), corals and macroalgae to turf (Shikine Island, Japan) (Inoue et al., 2013; Hall-Spencer et al., 2008; Agostini et al., 2018). Many of these shifts occurred at *p*CO₂ conditions projected by the mid-century and suggest that thresholds of tipping points may be reached in the near future. We project that in the future, many ecosystems (but not all) will shift from states dominated by the cover of marine forests or corals to more depauperate states.



280 The changes in the extent and structural complexity of biogenic habitat observed within CO₂ seeps can additionally mediate further biodiversity shifts, with the impact of ocean acidification on habitat-forming species projected to lead to lower associated species diversity (Sunday et al., 2016). Field surveys in a temperate Pacific CO₂ seep found that the ocean acidification-driven transition in habitat resulted in a reduction in the diversity of associated fish species, this loss is largely attributed to the disappearance of species that are closely associated with habitats that have undergone significant loss themselves. Moreover, it led to the selection of fish species that are more adapted to simplified ecosystems dominated by algae (Cattano et al., 2016). Similar declines in reef-associated macroinvertebrate communities, as well as small cryptic invertebrates, have also been observed in tropical CO₂ seeps (Fabricius et al., 2014; Plaisance et al., 2021).

285 Habitats play a crucial role in shaping the diversity of associated organisms, including macroinvertebrates and fishes; however, it is important to note that these organisms themselves also exert control over the habitats they inhabit through their grazing. In acidified conditions, it has been suggested that the top-down control exerted by some grazers may diminish, as evidenced by reductions in the diversity, abundance and size of many marine fauna observed at CO₂ seeps (Garilli et al., 2015; Harvey et al., 2018; Harvey et al., 2016). For example, there is a decrease in the number of sea urchin feeding halos at a Mediterranean CO₂ seeps (Kroeker et al., 2013a). Top-down control in ecological systems functions as a feedback loop, where the abundance and behaviour of grazers play a crucial role in shaping the population dynamics and composition of the habitat. With any loss or reduction of grazers and their top-down control, there is a potential for significant alterations in the overall structure and functioning of an ecosystem, and so diminished top-down control is likely contributing towards the simplification of coastal ecosystems.

300 The early-stage recruitment and trajectory of community development are important mechanisms responsible for altering shallow marine communities exposed to ocean acidification. Ocean acidification has notable effects on the composition of prokaryotic biofilm communities on both natural and artificial substrates deployed in CO₂ seeps (Kerfahi et al., 2014; Taylor et al., 2014). Similarly, studies on eukaryotic biofilm communities in these seep environments have also observed changes in community composition (Allen et al., 2021). Collectively, these findings indicate that the ecological patterns driving biofilm community responses in CO₂ seeps are characterised by a selection for distinct cohorts of organisms, where the conditions at the reference and elevated pCO₂ sites represent distinct niches. When examining the longer-term development of communities, the divergence of community composition over time is often influenced by changes in competitive interactions among habitat-forming organisms (Kroeker et al., 2013a; Kroeker et al., 2011). In general, dominant species tend to outcompete others during the early stages of succession, rapidly outgrowing or overgrowing them, which can result in the community being locked into a depauperate and low-complexity state.



When examining the impact of ocean acidification on ecosystem change, the interplay of physiological thresholds and ecological interactions, such as habitat provisioning, community development, and top-down control, becomes crucial. The absence or loss of certain species indicates that either their physiology and/or their ecological dynamics are unable to cope with the adverse effects of ocean acidification. For example, even if a species has the physiological capacity to tolerate ocean acidification conditions, it may still be competitively excluded during community development, leading to its absence in elevated $p\text{CO}_2$ conditions. For example, if crucial settlement substrates such as coralline algae are absent, this ocean acidification may then have large indirect negative impacts on corals, irrespective of direct effects (Fabricius et al., 2015; Fabricius et al., 2011; Fabricius et al., 2017). However, our understanding of the physiological mechanisms that drive these changes are poorly understood in most cases, and measurements of direct competition are often lacking in both *in situ* and laboratory research. Individual CO_2 seeps, while analogues for specific factors like CO_2 , may not fully represent the broader influences of climate change. Subsequently, in recent years, research has moved towards replicating research across multiple CO_2 seeps (to encompass multiple environmental drivers alongside ocean acidification) (Cornwall et al., 2017b; Comeau et al., 2022), as well as making use of other natural analogues like semi-enclosed lagoons (e.g., Palau and New Caledonia), which have provided important insights into corals that have adapted to these systems (Kurihara et al., 2021; Tanvet et al., 2023).

4. Conclusions

Ocean acidification has complex biological consequences, affecting calcification, photo-physiology, and pH regulation in various marine taxa. These direct impacts have been quantified in more than one thousand research articles to date. However, more research is required on internal pH regulation of various taxa (see Table 1), and up to date meta-analyses of non-calcareous taxa (see table 2) is also required. Quantitative projections of the tipping points at which CO_2 will have negative (or positive) impacts is also required for most taxa, where here we generally rely on semi-qualitative assessments for all taxa except corals and coralline algae. These negative and positive impacts will have flow on effects at ecological levels. Natural analogues, like CO_2 seeps, offer insights into the long-term effects of ocean acidification on marine ecosystems. These natural laboratories show shifts in habitat structure and community composition along $p\text{CO}_2$ gradients, leading to potential biodiversity loss and ecosystem simplification. Overall, ocean acidification has complex and far-reaching effects on marine life, and predicting specific tipping points for different taxa is challenging due to the multitude of factors involved. Continued research and monitoring efforts are essential to comprehend and address the impacts of ocean acidification on marine ecosystems in the future.

335 Acknowledgements



CEC was funded by a Rutherford Discovery Fellowship from Te Apārangi The Royal Society of New Zealand (VUW 1701).

Conflicts of interest

The contact author has declared that none of the authors has any competing interests.

References

- 340 Agostini, S., Harvey, B. P., Wada, S., Kon, K., Milazzo, M., Inaba, K., and Hall-Spencer, J. M.: Ocean acidification drives community shifts towards simplified non-calcified habitats in a subtropical–temperate transition zone, *Scientific Reports*, 8, 11354, 10.1038/s41598-018-29251-7, 2018.
- Albright, R., Takeshita, Y., Koweek, D. A., Ninokawa, A., Wolfe, K., Rivlin, T., Nebuchina, Y., Young, J., and Caldeira, K.: Carbon dioxide addition to coral reef waters suppresses net community calcification, *Nature*, 555, 516, 10.1038/nature25968
- 345 <https://www.nature.com/articles/nature25968#supplementary-information>, 2018.
- Albright, R., Caldeira, L., Hosfelt, J., Kwiatkowski, L., Maclaren, J. K., Mason, B. M., Nebuchina, Y., Ninokawa, A., Pongratz, J., Ricke, K. L., Rivlin, T., Schneider, K., Sesboüé, M., Shamberger, K., Silverman, J., Wolfe, K., Zhu, K., and Caldeira, K.: Reversal of ocean acidification enhances net coral reef calcification, *Nature*, 531, 362, 10.1038/nature17155
- <https://www.nature.com/articles/nature17155#supplementary-information>, 2016.
- 350 Allemand, D., Ferrier-Pagès, C., Furla, P., Houlbrèque, F., Puverel, S., Reynaud, S., Tambutte, E., Tambutte, S., and Zoccola, D.: Biomineralisation in reef-building corals: from molecular mechanisms to environmental control, *C.R. Palevol.*, 3, 453-467, 2004.
- Allen, R. J., Summerfield, T. C., Harvey, B. P., Agostini, S., Rastrick, S. P., Hall-Spencer, J. M., and Hoffmann, L. J.: Species turnover underpins the effect of elevated CO₂ on biofilm communities through early succession, *Climate Change Ecology*, 2, 100017, 2021.
- Andersson, A. J. and Gledhill, D. K.: Ocean acidification and coral reefs: effects on breakdown, dissolution, and net ecosystem calcification, *Annual Review of Marine Science*, 5, 1-28, 2013.
- 355 Andersson, A. J., Mackenzie, F. T., and Bates, N. R.: Life on the margin: implications of ocean acidification on Mg-calcite, high latitude and cold-water marine calcifiers, *Marine Ecology Progress Series*, 373, 265-273, 2008.
- Anthony, K. R. N., Kline, D. I., Diaz-Pulido, G., Dove, S., and Hoegh-Guldberg, O.: Ocean acidification causes bleaching and productivity loss in coral reef builders, *Proceedings of the National Academy of Sciences*, 105, 17442-17446, 2008.
- 360 Baggini, C., Salomidi, M., Voutsinas, E., Bray, L., Krasakopoulou, E., and Hall-Spencer, J.: Seasonality affects macroalgal community response to increases in pCO₂, *PLoS ONE*, 9, e106520, 2014.
- Bathurst, R. G.: *Carbonate sediments and their diagenesis*, Elsevier 1972.
- Blain, C. O., Kulins, S., Radford, C. A., Sewell, M. A., and Shears, N. T.: Heterogeneity around CO₂ vents obscures the effects of ocean acidification on shallow reef communities, *ICES Journal of Marine Science*, 78, 3162-3175, 10.1093/icesjms/fsab184, 2021.
- 365 Carter, H. A., Ceballos-Osuna, L., Miller, N. A., and Stillman, J. H.: Impact of ocean acidification on metabolism and energetics during early life stages of the intertidal porcelain crab *Petrolisthes cinctipes*, *Journal of Experimental Biology*, 216, 1412-1422, 10.1242/jeb.078162, 2013.
- Cattano, C., Giomi, F., and Milazzo, M.: Effects of ocean acidification on embryonic respiration and development of a temperate wrasse living along a natural CO₂ gradient, *Conservation Physiology*, 4, 10.1093/conphys/cov073, 2016.
- 370 Chan, N. C. S. and Connolly, S. R.: Sensitivity of coral calcification to ocean acidification: A meta-analysis, *Global Change Biology*, 19, 282-290, 2013.
- Comeau, S., Carpenter, R. C., and Edmunds, P. J.: Coral reef calcifiers buffer their responses to ocean acidification using both bicarbonate and carbonate, *Proceedings of the Royal Society B: Biological Sciences*, 280, doi: 10.1098/rspb.2012.2374, 2013a.
- Comeau, S., Edmunds, P. J., Spindel, N. B., and Carpenter, R. C.: The responses of eight coral reef calcifiers to increasing partial pressure of CO₂ do not exhibit a tipping point, *Limnology and Oceanography*, 58, 388-398, 2013b.
- 375 Comeau, S., Edmunds, P. J., Spindel, N. B., and Carpenter, R. C.: Fast coral reef calcifiers are more sensitive to ocean acidification in short-term laboratory incubations, *Limnology and Oceanography*, 59, 1081-1091, doi:10.4319/lo.2014.59.3.1081, 2014.
- Comeau, S., Cornwall, C. E., DeCarlo, T. M., Doo, S. S., Carpenter, R. C., and McCulloch, M. T.: Resistance to ocean acidification in coral reef taxa is not gained by acclimatization, *Nature Climate Change*, 9, 477-483, 10.1038/s41558-019-0486-9, 2019.



- 380 Comeau, S., Cornwall, C. E., Shlesinger, T., Hoogenboom, M., Mana, R., McCulloch, M. T., and Rodolfo-Metalpa, R.: pH variability at volcanic CO₂ seeps regulates coral calcifying fluid chemistry, *Global Change Biology*, 28, 2751-2763, <https://doi.org/10.1111/gcb.16093>, 2022.
Cornwall, C. E., Comeau, S., and McCulloch, M. T.: Coralline algae elevate pH at the site of calcification under ocean acidification, *Global Change Biology*, 23, 4245-4256, 2017a.
- 385 Cornwall, C. E., Comeau, S., DeCarlo, T. M., Moore, B., D'Alexis, Q., and McCulloch, M. T.: Resistance of corals and coralline algae to ocean acidification: physiological control of calcification under natural pH variability, *Proceedings of the Royal Society B: Biological Sciences*, 285, 10.1098/rspb.2018.1168, 2018.
Cornwall, C. E., Reville, A. T., Hall-Spencer, J. M., Milazzo, M., Raven, J. A., and Hurd, C. L.: Inorganic carbon physiology underpins macroalgal responses to elevated CO₂, *Scientific Reports*, 7, 46297, 10.1038/srep46297, 2017b.
- 390 Cornwall, C. E., Harvey, B. P., Comeau, S., Cornwall, D. L., Hall-Spencer, J. M., Peña, V., Wada, S., and Porzio, L.: Understanding coralline algal responses to ocean acidification: Meta-analysis and synthesis, *Global Change Biology*, 28, 362-374, <https://doi.org/10.1111/gcb.15899>, 2022.
Cornwall, C. E., Comeau, S., DeCarlo, T. M., Larcombe, E., Moore, B., Giltrow, K., Puerzer, F., D'Alexis, Q., and McCulloch, M. T.: A coralline alga gains tolerance to ocean acidification over multiple generations of exposure, *Nature Climate Change*, 10, 143-146, 10.1038/s41558-019-0681-8, 2020.
- 395 Cornwall, C. E., Carlot, J., Branson, O., Courtney, T. A., Harvey, B. P., Perry, C. T., Andersson, A. J., Diaz-Pulido, G., Johnson, M. D., Kennedy, E., Krieger, E. C., Mallela, J., McCoy, S. J., Nugues, M. M., Quinter, E., Ross, C. L., Ryan, E., Saderne, V., and Comeau, S.: Crustose coralline algae can contribute more than corals to coral reef carbonate production, *Communications Earth & Environment*, 4, 105, 10.1038/s43247-023-00766-w, 2023.
- 400 Cornwall, C. E., Comeau, S., Kornder, N. A., Perry, C. T., van Hooideonk, R., DeCarlo, T. M., Pratchett, M. S., Anderson, K. D., Browne, N., Carpenter, R., Diaz-Pulido, G., D'Olivo, J. P., Doo, S. S., Figueiredo, J., Fortunato, S. A. V., Kennedy, E., Lantz, C. A., McCulloch, M. T., González-Rivero, M., Schoepf, V., Smithers, S. G., and Lowe, R. J.: Global declines in coral reef calcium carbonate production under ocean acidification and warming, *Proceedings of the National Academy of Sciences*, 118, e2015265118, 10.1073/pnas.2015265118, 2021.
DeCarlo, T. M., Comeau, S., Cornwall, C. E., and McCulloch, M. T.: Coral resistance to ocean acidification linked to increased calcium at the site of calcification, *Proceedings of the Royal Society B: Biological Sciences*, 285, 20180564, 10.1098/rspb.2018.0564, 2018.
- 405 Doney, S. C., Fabry, V. J., Feely, R. A., and Kleypas, J. A.: Ocean acidification: the other CO₂ problem, *Annual Review of Marine Science*, 1, 169-192, 2009.
Drake, J. L., Mass, T., Haramaty, L., Zelzion, E., Bhattacharya, D., and Falkowski, P. G.: Proteomic analysis of skeletal organic matrix from the stony coral *Stylophora pistillata*, *Proceedings of the National Academy of Sciences*, 110, 3788-3793, doi:10.1073/pnas.1301419110, 2013.
- 410 Enochs, I. C., Manzello, D. P., Donham, E. M., Kolodziej, G., Okano, R., Johnston, L., Young, C., Iguel, J., Edwards, C. B., Fox, M. D., Valentino, L., Johnson, S., Benavente, D., Clark, S. J., Carlton, R., Burton, T., Eynaud, Y., and Price, N. N.: Shift from coral to macroalgae dominance on a volcanically acidified reef, *Nature Climate Change*, 5, 1083-1088, 10.1038/nclimate2758, 2015.
Fabricius, K., Klübenschedl, A., Harrington, L., Noonan, S., and De'Ath, G.: In situ changes of tropical crustose coralline algae along carbon dioxide gradients, *Scientific reports*, 5, 9537, 10.1038/srep09537, 2015.
- 415 Fabricius, K., Langdon, C., Uthicke, S., Humphrey, C., Noonan, S., De'ath, G., Okazaki, R., Muehllehner, N., Glas, M. S., and Lough, J. M.: Losers and winners in coral reefs acclimatized to elevated carbon dioxide concentrations, *Nature Climate Change*, 1, 165-169, 2011.
Fabricius, K. E., De'ath, G., Noonan, S., and Uthicke, S.: Ecological effects of ocean acidification and habitat complexity on reef-associated macroinvertebrate communities, *Proceedings of the Royal Society B*, 281, 20132479, 2014.
- 420 Fabricius, K. E., Noonan, S. H. C., Abrego, D., Harrington, L., and De'ath, G.: Low recruitment due to altered settlement substrata as primary constraint for coral communities under ocean acidification, *Proceedings of the Royal Society B: Biological Sciences*, 284, 20171536, 10.1098/rspb.2017.1536, 2017.
Falkenberg, L. J., Styan, C. A., and Havenhand, J. N.: Sperm motility of oysters from distinct populations differs in response to ocean acidification and freshening, *Scientific Reports*, 9, 7970, 10.1038/s41598-019-44321-0, 2019.
- 425 Feely, R. A., Sabine, C. L., Hernandez-Ayon, J. M., Ianson, D., and Hales, B.: Evidence for upwelling of corrosive "acidified" water onto the coastal shelf, *Science*, 320, 1490-1492, 2008.
Feely, R. A., Sabine, C. L., Lee, K., Berelson, W., Kleypas, J. A., Fabry, V. J., and Millero, F. J.: Impact of anthropogenic CO₂ on the CaCO₃ system in the oceans, *Science*, 305, 362-366, 2004.
- 430 Garilli, V., Rodolfo-Metalpa, R., Scuderi, D., Brusca, L., Parrinello, D., Rastrick, S. P. S., Foggo, A., Twitchett, R. J., Hall-Spencer, J. M., and Milazzo, M.: Physiological advantages of dwarfing in surviving extinctions in high-CO₂ oceans, *Nature Climate Change*, 5, 678-682, 10.1038/nclimate2616, 2015.
Gazeau, F., Parker, L. M., Comeau, S., Gattuso, J.-P., O'Connor, W. A., Martin, S., Pörtner, H.-O., and Ross, P. M.: Impacts of ocean acidification on marine shelled molluscs, *Marine Biology*, 160, 2207-2245, 10.1007/s00227-013-2219-3, 2013.
- 435 Glynn, P. W.: Eastern Pacific Coral Reef Ecosystems, in: *Coastal Marine Ecosystems of Latin America*, edited by: Seeliger, U., and Kjerfve, B., Springer Berlin Heidelberg, Berlin, Heidelberg, 281-305, 10.1007/978-3-662-04482-7_20, 2001.



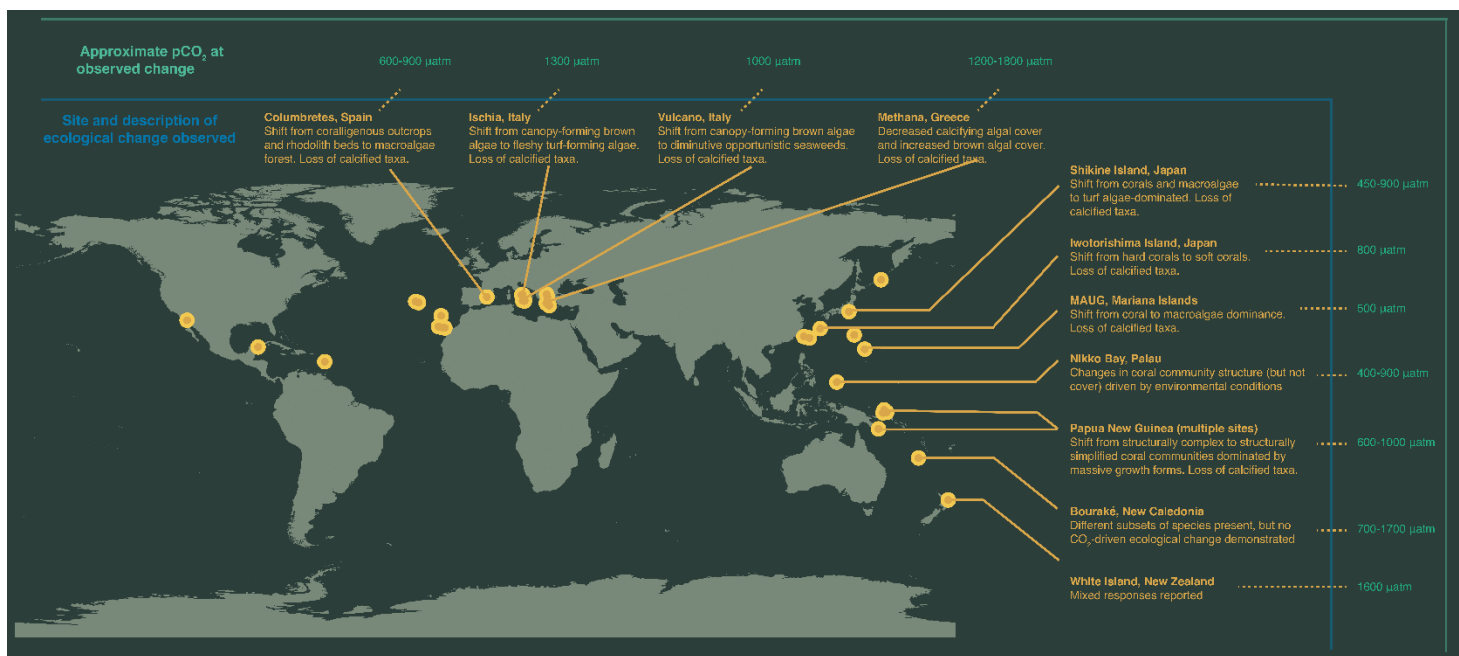
- Hall-Spencer, J. M., Rodolfo-Metalpa, R., Martin, S., Ransome, E., Fine, M., Turner, S. M., Rowley, S. J., Tedesco, D., and Buia, M. C.: Volcanic carbon dioxide vents show ecosystem effects of ocean acidification, *Nature*, 454, 96-99, 2008.
- Harvey, B. P., Gwynn-Jones, D., and Moore, P. J.: Meta-analysis reveals complex marine biological responses to the interactive effects of ocean acidification and warming, *Ecology and evolution*, 3, 1016-1030, 2013.
- 440 Harvey, B. P., Agostini, S., Wada, S., Inaba, K., and Hall-Spencer, J. M.: Dissolution: The Achilles' Heel of the Triton Shell in an Acidifying Ocean, *Frontiers in Marine Science*, 5, 10.3389/fmars.2018.00371, 2018.
- Harvey, B. P., McKeown, N. J., Rastrick, S. P. S., Bertolini, C., Foggo, A., Graham, H., Hall-Spencer, J. M., Milazzo, M., Shaw, P. W., Small, D. P., and Moore, P. J.: Individual and population-level responses to ocean acidification, *Scientific Reports*, 6, 20194, 10.1038/srep20194, 2016.
- 445 Hendriks, I. E., Duarte, C. M., and Álvarez, M.: Vulnerability of marine biodiversity to ocean acidification: a meta-analysis, *Estuar. Coast. Shelf Sci.*, 86, 157-164, 2010.
- Hepburn, C. D., Pritchard, D. W., Cornwall, C. E., McLeod, R. J., Beardall, J., Raven, J. A., and Hurd, C. L.: Diversity of Carbon use strategies in a kelp forest community: implications for a high CO₂ ocean, *Global Change Biology*, 17, 2488-2497, 2011.
- Hurd, C. L., Beardall, J., Comeau, S., Cornwall, C. E., Havenhand, J. N., Munday, P. L., Parker, L. M., Raven, J. A., and McGraw, C. M.: Ocean acidification as a multiple driver: how interactions between changing seawater carbonate parameters affect marine life, *Marine and Freshwater Research*, -, <https://doi.org/10.1071/MF19267>, 2019.
- 450 Inoue, S., Kayanne, H., Yamamoto, S., and Kurihara, H.: Spatial community shift from hard to soft corals in acidified water, *Nature Climate Change*, 3, 683-687, 10.1038/nclimate1855, 2013.
- Jokiel, P. L.: Coral reef calcification: carbonate, bicarbonate and proton flux under conditions of increasing ocean acidification, *Proceedings of the Royal Society B: Biological Sciences*, 280, 1764, 2013.
- 455 Kerfahi, D., Hall-Spencer, J. M., Tripathi, B. M., Milazzo, M., Lee, J., and Adams, J. M.: Shallow water marine sediment bacterial community shifts along a natural CO₂ gradient in the Mediterranean Sea off Vulcano, Italy, *Microb Ecol*, 67, 819-828, 10.1007/s00248-014-0368-7, 2014.
- Knowlton, N.: Thresholds and Multiple Stable States in Coral Reef Community Dynamics, *American Zoologist*, 32, 674-682, 10.1093/icb/32.6.674, 1992.
- 460 Kornder, N. A., Riegl, B. M., and Figueiredo, J.: Thresholds and drivers of coral calcification responses to climate change, *Global Change Biology*, 24, 5084-5095, doi:10.1111/gcb.14431, 2018.
- Kroeker, K. J., Gambi, M. C., and Micheli, F.: Community dynamics and ecosystem simplification in a high-CO₂ ocean, *Proceedings of the National Academy of Science, USA*, 110, 12721-12726, 2013a.
- 465 Kroeker, K. J., Kordas, R. L., Crim, R. N., and Singh, G. G.: Meta-analysis reveals negative yet variable effects of ocean acidification on marine organisms, *Ecology letters*, 13, 1419-1434, 2010.
- Kroeker, K. J., Micheli, F., Gambi, M. C., and Martz, T. R.: Divergent ecosystem responses within a benthic marine community to ocean acidification, *Proc. Natl. Acad. Sci. U.S.A.*, 108, 14515-14520, 2011.
- 470 Kroeker, K. J., Kordas, R. L., Crim, R. N., Hendriks, I. E., Ramajo, L., Singh, G. G., Duarte, C. M., and Gattuso, J. P.: Impacts of ocean acidification on marine organisms: quantifying sensitivities and interaction with warming, *Global Change Biology*, 19, 1884-1896, 2013b.
- Kurihara, H., Watanabe, A., Tsugi, A., Mimura, I., Hongo, C., Kawai, T., Reimer, J. D., Kimoto, K., Gouezo, M., and Golbuu, Y.: Potential local adaptation of corals at acidified and warmed Nikko Bay, Palau, *Scientific Reports*, 11, 11192, 10.1038/s41598-021-90614-8, 2021.
- Leung, J. Y. S., Zhang, S., and Connell, S. D.: Is Ocean Acidification Really a Threat to Marine Calcifiers? A Systematic Review and Meta-Analysis of 980+ Studies Spanning Two Decades, *Small*, 18, 2107407, <https://doi.org/10.1002/sml.202107407>, 2022.
- 475 Linares, C., Vidal, M., Canals, M., Kersting, D. K., Amblas, D., Aspillaga, E., Cebrian, E., Delgado-Huertas, A., Diaz, D., Garrabou, J., Hereu, B., Navarro, L., Teixido, N., and Ballesteros, E.: Persistent natural acidification drives major distribution shifts in marine benthic ecosystems, *Proceedings of the Royal Society B-Biological Sciences*, 282, 7, 10.1098/rspb.2015.0587, 2015.
- Maberly, S. C., Berthelot, S. A., Stott, A. W., and Gontero, B.: Adaptation by macrophytes to inorganic carbon down river with naturally variable concentrations of CO₂, *Journal of Plant Physiology*, 172, 120-127, 2014.
- 480 Maggioni, F., Pujo-Pay, M., Aucan, J., Cerrano, C., Calcinai, B., Payri, C., Benzoni, F., Letourneur, Y., and Rodolfo-Metalpa, R.: The Bouraké semi-enclosed lagoon (New Caledonia) – a natural laboratory to study the lifelong adaptation of a coral reef ecosystem to extreme environmental conditions, *Biogeosciences*, 18, 5117-5140, 10.5194/bg-18-5117-2021, 2021.
- Manzello, D. P.: Ocean acidification hot spots: spatiotemporal dynamics of the seawater CO₂ system of eastern Pacific coral reefs, *Limnol. Oceanogr.*, 55, 239-248, 2010.
- 485 Manzello, D. P., Kleypas, J. A., Budd, D. A., Eakin, C. M., Glynn, P. W., and Langdon, C.: Poorly cemented coral reefs of the eastern tropical Pacific: Possible insights into reef development in a high-CO₂ world, *Proceedings of the National Academy of Sciences*, 105, 10450, 10.1073/pnas.0712167105, 2008.
- Marin, F., Luquet, G., Marie, B., and Medakovic, D.: Molluscan Shell Proteins: Primary Structure, Origin, and Evolution, in: *Current Topics in Developmental Biology*, Academic Press, 209-276, [https://doi.org/10.1016/S0070-2153\(07\)80006-8](https://doi.org/10.1016/S0070-2153(07)80006-8), 2007.



- 490 McCulloch, M., Trotter, J., Montagna, P., Falter, J., Dunbar, R., Freiwald, A., Försterra, G., Correa, M. L., Maier, C., Rüggeberg, A., and Taviani, M.: Resilience of cold-water scleratinian corals to ocean acidification: boron isotopic systematics of pH and saturation state up-regulation, *Geochemica et Cosmochimica Acta*, 87, 21-34, 2012a.
McCulloch, M. T., Falter, J., Trotter, J., and Montagna, P.: Coral resilience to ocean acidification and global warming through pH up-regulation, *Nature Climate Change*, 2, 623-627, 2012b.
- 495 McCulloch, M. T., D'Olivo, J. P., Falter, J., Holcomb, M., and Trotter, J. A.: Coral calcification in a changing world and the interactive dynamics of pH and DIC upregulation, *Nature Communications*, 8, 15686, 2017.
Meyer, J. and Riebesell, U.: Reviews and Syntheses: Responses of coccolithophores to ocean acidification: a meta-analysis, *Biogeosciences*, 12, 1671-1682, 10.5194/bg-12-1671-2015, 2015.
Moore, B., Comeau, S., Bekaert, M., Cossais, A., Purdy, A., Larcombe, E., Puerzer, F., McCulloch, M. T., and Cornwall, C. E.: Rapid multi-generational acclimation of coralline algal reproductive structures to ocean acidification, *Proceedings of the Royal Society B: Biological Sciences*, 288, 20210130, doi:10.1098/rspb.2021.0130, 2021.
- 500 Morse, J. W. and MacKenzie, F. T.: *Geochemistry of Sedimentary Carbonates*, 1990.
Moy, F. E. and Christie, H.: Large-scale shift from sugar kelp (*Saccharina latissima*) to ephemeral algae along the south and west coast of Norway, *Marine Biology Research*, 8, 309-321, 10.1080/17451000.2011.637561, 2012.
- 505 Nagelkerken, I., Russell, B. D., Gillanders, B. M., and Connell, S. D.: Ocean acidification alters fish populations indirectly through habitat modification, *Nature Climate Change*, 6, 89-93, 10.1038/nclimate2757, 2016.
Okazaki, R. R., Towle, E. K., van Hooijdonk, R., Mor, C., Winter, R. N., Piggot, A. M., Cuning, R., Baker, A. C., Klaus, J. S., Swart, P. K., and Langdon, C.: Species-specific responses to climate change and community composition determine future calcification rates of Florida Keys reefs, *Global Change Biology*, 23, 1023-1035, 10.1111/gcb.13481, 2017.
- 510 Parker, L. M., Scanes, E., O'Connor, W. A., and Ross, P. M.: Transgenerational plasticity responses of oysters to ocean acidification differ with habitat, *Journal of Experimental Biology*, 224, 10.1242/jeb.239269, 2021.
Parker, L. M., Ross, P. M., #039, Connor, W. A., Pörtner, H. O., Scanes, E., and Wright, J. M.: Predicting the Response of Molluscs to the Impact of Ocean Acidification, *Biology*, 2, 651-692, 2013.
Plaisance, L., Matterson, K., Fabricius, K., Drovetski, S., Meyer, C., and Knowlton, N.: Effects of low pH on the coral reef cryptic invertebrate communities near CO₂ vents in Papua New Guinea, *PLOS ONE*, 16, e0258725, 10.1371/journal.pone.0258725, 2021.
- 515 Porzio, L., Buia, M. C., and Hall-Spencer, J. M.: Effects of ocean acidification on macroalgal communities, *Journal of Experimental Marine Biology and Ecology*, 400, 278-287, 2011.
Ramajo, L., Pérez-León, E., Hendriks, I. E., Marbà, N., Krause-Jensen, D., Sejr, M. K., Blicher, M. E., Lagos, N. A., Olsen, Y. S., and Duarte, C. M.: Food supply confers calcifiers resistance to ocean acidification, *Scientific reports*, 6, 19374, 2016.
- 520 Raven, J. A. and Beardall, J.: CO₂ concentrating mechanisms and environmental change, *Aquatic Botany*, 118, 24-37, 2014.
Raven, J. A. and Beardall, J.: The ins and outs of CO₂, *Journal of Experimental Botany*, 67, 1-13, 2016.
Raven, J. A., Cockell, C. S., and De La Rocha, C. L.: The evolution of inorganic carbon concentrating mechanisms in photosynthesis, *Phil. Trans. R. Soc. B*, 363, 2641-2650, 2008.
Raven, J. A., Suggett, D. J., and Giordano, M.: Inorganic carbon concentrating mechanisms in free-living and symbiotic dinoflagellates and chromerids, *Journal of Phycology*, 56, 1377-1397, <https://doi.org/10.1111/jpy.13050>, 2020.
- 525 Raven, J. A., Giordano, M., Beardall, J., and Maberly, S. C.: Algal and aquatic plant carbon concentrating mechanisms in relation to environmental change, *Photosynth. Res.*, 109, 281-296, 2011.
Raven, J. A., Giordano, M., Beardall, J., and Maberly, S. C.: Algal evolution in relation to atmospheric CO₂: carboxylases, carbon-concentrating mechanisms and carbon oxidation, *Philosophical Transactions of the Royal Society B: Biological Sciences*, 367, 493-507, 2012.
- 530 Raven, J. A., Ball, L. A., Beardall, J., Giordano, M., and Maberly, S. C.: Algae lacking carbon-concentrating mechanisms, *Canadian Journal of Botany*, 83, 879-890, 10.1139/b05-074, 2005.
Raven, J. A., Johnston, A. M., Kübler, J. E., Korb, R. E., McInroy, S. G., Handley, L. L., Scrimgeour, C. M., Walker, D. I., Beardall, J., Vanderklift, M. A., Fredriksen, S., and Dunton, K. H.: Mechanistic interpretation of carbon isotope discrimination by marine macroalgae and seagrasses, *Funct. Plant Biol.*, 29, 355-378, 2002a.
- 535 Raven, J. A., Johnston, A. M., Kübler, J. E., Korb, R., McInroy, S. G., Handley, L. L., Scrimgeour, C. M., Walker, D. I., Beardall, J., Clayton, M. N., Vanderklift, M. A., Fredriksen, S., and Dunton, K. H.: Seaweeds in cold seas: evolution and carbon acquisition, *Ann. Bot.*, 90, 525-536, 2002b.
- 540 Ries, J. B.: Acid ocean cover up, *Nature Climate Change*, 1, 294-295, 2011.
Ries, J. B., Cohen, A. L., and McCorkle, D. C.: Marine calcifiers exhibit mixed responses to CO₂-induced ocean acidification, *Geology*, 37, 1131-1134, 2009.
Rivest, E. B., Comeau, S., and Cornwall, C. E.: The role of natural variability in shaping the response of coral reef organisms to climate change, *Current Climate Change Reports*, 3, 271-281, 2017.
Roberts, R. D.: A review of settlement cues for larval abalone (*Haliotis* spp.), *J. Shell. Res.*, 20, 571-586, 2001.



- 545 Schneider, K. and Erez, J.: The effect of carbonate chemistry on calcification and photosynthesis in the hermatypic coral *Acropora eurystoma*, *Limnology and Oceanography*, 51, 1284-1293, 2006.
- Schoepf, V., Carrion, S. A., Pfeifer, S. M., Naugle, M., Dugal, L., Bruyn, J., and McCulloch, M. T.: Stress-resistant corals may not acclimatize to ocean warming but maintain heat tolerance under cooler temperatures, *Nature Communications*, 10, 4031, 10.1038/s41467-019-12065-0, 2019.
- 550 Schubert, N., Alvarez-Filip, L., and Hofmann, L. C.: Systematic review and meta-analysis of ocean acidification effects in Halimeda: Implications for algal carbonate production, *Climate Change Ecology*, 4, 100059, <https://doi.org/10.1016/j.ecochg.2022.100059>, 2023.
- Stumpp, M., Hu, M. Y., Melzner, F., Gutowska, M. A., Dorey, N., Himmerkus, N., Holtmann, W. C., Dupont, S. T., Thorndyke, M. C., and Bleich, M.: Acidified seawater impacts sea urchin larvae pH regulatory systems relevant for calcification, *Proceedings of the National Academy of Sciences of the United States of America*, 109, 18192-18197, 10.1073/pnas.1209174109, 2012.
- 555 Sunday, J. M., Fabricius, K. E., Kroeker, K. J., Anderson, K. M., Brown, N. E., Barry, J. P., Connell, S. D., Dupont, S., Gaylord, B., Hall-Spencer, J. M., Klinger, T., Milazzo, M., Munday, P. L., Russell, B. D., Sanford, E., Thiyagarajan, V., Vaughan, M. L. H., Widdicombe, S., and Harley, C. D. G.: Ocean acidification can mediate biodiversity shifts by changing biogenic habitat, *Nature Climate Change*, 7, 81, 10.1038/nclimate3161, 2016.
- Tanvet, C., Camp, E. F., Sutton, J., Houlbrèque, F., Thouzeau, G., and Rodolfo-Metalpa, R.: Corals adapted to extreme and fluctuating seawater pH increase calcification rates and have unique symbiont communities, *Ecol Evol*, 13, e10099, 10.1002/ece3.10099, 2023.
- 560 Taylor, J. D., Ellis, R., Milazzo, M., Hall-Spencer, J. M., and Cunliffe, M.: Intertidal epilithic bacteria diversity changes along a naturally occurring carbon dioxide and pH gradient, *FEMS Microbiology Ecology*, 89, 670-678, <https://doi.org/10.1111/1574-6941.12368>, 2014.
- Thomsen, J. and Melzner, F.: Moderate seawater acidification does not elicit long-term metabolic depression in the blue mussel *Mytilus edulis*, *Mar. Biol.*, 157, 2667-2676, 2010.
- 565 Venn, A. A., Bernardet, C., Chabenat, A., Tambutté, E., and Tambutté, S.: Paracellular transport to the coral calcifying medium: effects of environmental parameters, *The Journal of Experimental Biology*, 223, jeb227074, 10.1242/jeb.227074, 2020.
- Venn, A. A., Tambutté, E., Holcomb, M., Laurent, J., Allemand, D., and Tambutté, S.: Impact of seawater acidification on pH at the tissue-skeleton interface and calcification in reef corals, *Proceedings of the National Academy of Sciences of the United States of America*, 110, 1634-1639, 10.1073/pnas.1216153110, 2013.
- 570 Wernberg, T., Bennett, S., Babcock, R. C., de Bettignies, T., Cure, K., Depczynski, M., Dufois, F., Fromont, J., Fulton, C. J., and Hovey, R. K.: Climate-driven regime shift of a temperate marine ecosystem, *Science*, 353, 169-172, 2016.
- Wittmann, A. C. and Pörtner, H. O.: Sensitivities of extant animal taxa to ocean acidification, *Nature Climate Change*, 3, 995-1001, 2013.



575



580 **Figure 1: Map of locations of high CO₂ locations used to make inferences regarding impacts of OA on future marine ecosystems.** Labeled are sites where patterns in ecosystem change is described at Columbretes, Spain (Linares et al., 2015); Ischia, Italy (Hall-Spencer et al., 2008; Porzio et al., 2011); Vulcano, Italy (Cornwall et al., 2017b); Methana, Greece (Baggini et al., 2014); Shikine Island, Japan (Agostini et al., 2018); Iwotorishima Island, Japan (Inoue et al., 2013); Maug, Mariana Islands (Enochs et al., 2015); Papua New Guinea (Fabricius et al., 2011; Comeau et al., 2022); Bouraké, New Caledonia (Maggioni et al., 2021); White Island, New Zealand (Nagelkerken et al., 2016; Blain et al., 2021).



585 Table 1: Summary of the impacts of ocean acidification on calcification/growth, photosynthetic rates and internal pH regulation, with tipping points from literature identified where possible. Top to bottom rows, taxa are listed from first to be negatively impacted at the top to most positively impacted at the bottom. Red = significant decrease in values from zero, yellow = no detectable difference, green = significant increase in values from zero, grey = not enough data to determine. Where possible, quantitative or qualitative tipping points given in $\mu\text{atm CO}_2$. Empty cells = not applicable. See Table 2 for further details on references used to generate tipping points.

Taxon	Calcification/growth	Photosynthetic rates	Internal pH regulation	References
Coralline algae	~538		~538 to 930	(Cornwall et al., 2021; Cornwall et al., 2022; Leung et al., 2022)
Corals	~572	~538 to 930	~538 to 930	(Leung et al., 2022; Cornwall et al., 2021)
Molluscs	~781			(Leung et al., 2022; Wittmann and Pörtner, 2013)
Sea urchins	~870			(Leung et al., 2022; Wittmann and Pörtner, 2013)
Polychaetes	~538 to 930			(Leung et al., 2022)
Foraminifera	~538 to 930			(Leung et al., 2022)
Coccolithophores	~538 to 930	~538 to 930		(Leung et al., 2022; Meyer and Riebesell, 2015)
Sponges				(Leung et al., 2022)



Echinoderms (non-sea urchin)			~538 to 930	(Leung et al., 2022)
<i>Halimeda</i> spp.				(Schubert et al., 2023)
Bryozoans				(Leung et al., 2022)
Crustaceans				(Leung et al., 2022)
Seagrasses		~538 to 930		(Kroeker et al., 2013b; Harvey et al., 2013)
Cephalopods	~538 to 930			(Leung et al., 2022)
Fish	~538 to 930			(Kroeker et al., 2013b; Harvey et al., 2013)
Diatoms	~538 to 930			(Kroeker et al., 2013b)
Non-calcareous seaweeds	~538 to 930			(Kroeker et al., 2013b; Harvey et al., 2013)

590



595

Table 2: Meta-analyses or pseudo-meta-analyses that support the results of table 1. Note: some studies excluded here due to merging of different taxa, inclusion of too few studies, or assessing only specific processes in combination with ocean acidification (e.g., Rivest et al., 2017; Ramajo et al., 2016). Studies listed chronologically and greyed out if their results not used due to complete overlap with more recent research. Note: fish behaviour excluded from these analyses. Meta-analyses checked by searching “ocean acidification” AND “meta-analysis” on July 2023. All non-regional assessments including photosynthesis, calcification or internal pH included below.

Study	Limited to specific taxa?	Year literature search conducted	Number of studies used	Taxa used to create table 1
Leung (2022)	Calcifying taxa	2020	985	All calcifying taxa except <i>Halimeda</i> spp.
Cornwall et al. (2022)	Coralline algae	2021	64	Coralline algae
Schubert et al. (2023)	<i>Halimeda</i> spp.	Unknown	31	<i>Halimeda</i> spp.
Cornwall et al. (2021)	Coralline algae, corals, bioerosion, sediments	2021	98	Coralline algae and corals
Kornder et al. (2018)	Corals	2016	62	None – though this study was used extensively in Cornwall et al. (2021)
Meyer and Riebesell (2015)	Coccolithophores	Unknown but 2015 or before	33	Coccolithophores (however, they often compared against 280 ppm CO ₂)
Harvey et al. (2013)	Calcifying algae, corals, crustaceans, echinoderms, molluscs, phytoplankton, fish, non-calcareous algae, seagrass	2012	107	All non-calcareous taxa except diatoms
Kroeker et al. (2013b)	Calcifying algae, corals, coccolithophores,	2012	155	All non-calcareous taxa



	molluscs, echinoderms, crustaceans, fish, fleshy algae, seagrasses, diatoms			
Wittmann and Pörtner (2013)	Crustaceans, echinoderms, corals, fish	2012	167	Echinoderms and molluscs
Chan and Connolly (2013)	Corals	2011	25	None
Kroecker et al. (2010)	Calcifying algae, corals, coccolithophores, molluscs, echinoderms, crustaceans, fish, fleshy algae, seagrasses, diatoms	2010	139	None
Hendriks et al. (2010)	Bivalves, corals, coccolithophores, phytoplankton, cyanobacteria	2010	59	None