

Manuscript “Global climate change and the Baltic Sea ecosystem: direct and indirect effects on species, communities and ecosystem functioning”

by Markku Viitasalo and Erik Bonsdorff, submitted to Earth System Dynamics.

Author comments for reviewer no. 3 (esd-2021-73-RC3-supplement.pdf).

All author replies to reviewer comments in red font.

### Anonymous reviewer #3

#### ESD specific reviewer statements

**1. Does the paper address relevant scientific questions within the scope of ESD?**

Yes.

**2. Does the paper present novel concepts, ideas, tools, or data?**

Not yet so much. There are new tools (or methods) to better bridge gaps in cross-compartment community structure analyses (e.g. by fuzzy coding or metabarcoding). But there are also new methods (namely compound specific isotope analyses of amino acid nitrogen, CSIA) that bridge gaps in cross-compartment functional diversity analyses more directly and which, different to other “multi trait” approaches, include significant reduction of complexity by addressing functional groups (according to Tilman 2001: phototrophs, mixotrophs, heterotrophs, herbivores, different levels of carnivores,) rather than individual traits or taxonomic groups. CSIA allows for the direct measurement of the mean trophic position of a field sample as one key cross-compartment functional trait (according to Tilman 2001) and of the dominant inorganic nitrogen source used for growth from a field sample as second key cross-compartment functional trait (Tilman 2001) for end-to-end analyses (e.g., physics to fish to human sectors, in sensu Peck et al., 2018) of food webs. Both traits can directly be determined from a single field sample independent from the compartment it comes from (e.g. mixed phytoplankton, mixed mesozooplankton, bivalves, herring, cod, seagulls) and can directly be used as “common currency” across all compartments (e.g., physics to fish to human sectors, in sensu Peck et al., 2018) to calibrate and validate current biogeochemical models. For example, no biogeochemical model yet accounts for the mass and energy loss for fish that must be related to the shift in mean trophic position of mesozooplankton from herbivory (TP of 2) to carnivory (TP of 3) during cyanobacterial blooms in the central Baltic Sea (see specific comment #) and which can help to explain the loss in apex predators in the Baltic Sea.

Thank You for the critical comments, we have now revised the ms. thoroughly and used a more critical approach. For instance, we have included more scrutiny on the reviewed papers and their methods, included assessment of consensus and dissensus in Conclusions, and also added new text on the potential of novel methods in Knowledge gaps (see response to the Specific comment no. 79).

**3. Are substantial conclusions reached?**

Yes, but they are sometimes inconsistent with parts in the earlier text. For example, I still don't know if the research up to now points to N<sub>2</sub>-fixing, unpalatable cyanobacteria as “winner” of cc or not. So conclusions should be somewhat refined.

We agree that projecting structure and functioning of the pelagic ecosystem with 3D modelling in the distant future (e.g., for the year 2100) is challenging and involves many assumptions. In our review we note that several 3D modelling studies project (in a BAU scenario) an increase of Cyanobacteria in a warmer and more stratified Baltic Sea, but in the Knowledge gaps and

Conclusions try to highlight the associated uncertainties by noting the dissensus between modelling studies and certain short-term monoculture and mesocosm studies. We emphasize that also the modelling studies propose that successful reduction of nutrient loading from land (according to HELCOM BSAP) would outweigh the effects of proceeding climate change and decrease the cyanobacteria biomass, or at least diminish the occurrence of very large blooms.

**4. Are the scientific methods and assumptions valid and clearly outlined?**

Generally yes. Yet, I feel like important empirical field based studies are still missing in this review as marked in the specific comments.

Thank you for the comment. We have added the suggested references, as pointed out below.

**5. Are the results sufficient to support the interpretations and conclusions?**

See point 3.

Response in point 3.

**6. Is the description of experiments and calculations sufficiently complete and precise to allow their reproduction by fellow scientists (traceability of results)?**

Pls explain how you found and chose your reviewed literature (e.g. did you use google scholar, did you visit the websites of major Baltic Sea research institutes for most recent publications etc.?).

A new Chapter 2, Definitions and review methods, is now included in the ms. The main method was Web of Science and we only included papers published in 2010-2021 (the period since BACC II; see Chapter 22 for a more detailed explanation of the review method).

We did not specifically look for institute web pages. We want to emphasize that this review is not a full systematic review of all research done on climate change effects on the Baltic Sea. Rather, we highlight the variety of field, experimental and modelling studies on this subject and summarise what can be concluded from the evidence published since 2010.

**7. Do the authors give proper credit to related work and clearly indicate their own new/original contribution?**

Not applicable for an invited review article, I think.

Ok.

**8. Does the title clearly reflect the contents of the paper?**

Authors should definitely add a definition of ecosystem functioning into your introduction or as a glossary, e.g. Ecosystem Functioning (Tilman 2001): The rate, level, or temporal dynamics of one or more ecosystem processes like primary production or nutrient gain or loss.

A new Chapter 2, Definitions and review methods is now included and the definitions are given there. We apply Tilman's (2001) definition of ecosystem functioning.

**9. Does the abstract provide a concise and complete summary?**

I will suggest a major revision and there might be changes in the abstract necessary due to that. For example, what about the internal P storage in the sediments that gets released under anoxic conditions? How will that influence the projection of future cyanobacterial blooms in the Baltic Sea?

Abstract is now revised and adjusted according to the suggestions provided. A notion of the importance of internal loading of P is also there.

**10. Is the overall presentation well structured and clear?**

The authors could more systematically show, which direct and indirect effects have been addressed by which kind of research. Generally, I feel that new knowledge based on empirical research did not always find its way into here, if the “title” did not include certain key words. See also reply to 6.

We have now assessed the potential direct and indirect climate related effects on species and ecosystem functioning, and also made an attempt to the attribution to either global (anthropogenic) climate change and more natural variations in climate related parameters. We have added missing papers as suggested.

**11. Is the language fluent and precise?**

There are some minor spelling and grammar errors that need to be corrected for.

Language errors corrected as suggested.

**12. Are mathematical formulae, symbols, abbreviations, and units correctly defined and used?**

Not applicable.

Ok.

**13. Should any parts of the paper (text, formulae, figures, tables) be clarified, reduced, combined, or eliminated?**

The structure could be sharpened a little bit maybe. For example, I find it more intuitive to start with phototrophs like phytoplankton and cyanobacteria rather than with “Microbial communities”. Also, cyanobacteria is a very broad group. Maybe palatable cyanobacteria should be differentiated from unpalatable ones, as well as those that do fix nitrogen from those that don’t fix nitrogen but only profit from leaking out of diazotroph nitrogen from the N<sub>2</sub>-fixing ones. More in the revised pdf attached.

Thank you for the comment, we have now restructured the text. The order is now: (1) Phytoplankton (2) Cyanobacteria (now separated from “Phytoplankton and Cyanobacteria”), (3) Zooplankton, etc. The chapter Bacteria (instead of “Microbial communities”) is now presented last, after Fish, before the chapter Climate change and ecosystem structure and function.

We decided to move relevant parts of the chapter “Climate change and primary production” to the chapters “Phytoplankton” and “Cyanobacteria”, as there was much overlap between these three chapters.

We have also clarified the distinctions between palatable and less palatable cyanobacteria, and explained the N<sub>2</sub>-dynamics as presented in the responses to detailed comments, below.

**14. Are the number and quality of references appropriate?**

There are important papers missing at the moment, I think. Pls see the revised pdf for details.

Suggested papers, and some others, added.

**15. Is the amount and quality of supplementary material appropriate?**

Not applicable.

Ok.

## General comment

I enjoyed reading and reviewing this manuscript and I think it's worth publishing. That's said, I think that there are still many improvements possible to make it even better. For example, the authors did not find yet a good balance between a "too detailed" revision of a study and a "too brief" review of a study. In many places, I have a quarrel with the statements of many studies, which are often generalized beyond recognition (anytime I request: "Pls be more specific"). Maybe the authors can add some details here and there, namely where mechanisms are mentioned but which are hardly explained in sufficient detail (pointed out below in the specific comments).

The scientific community currently is struggling to find "the right" definition of functional diversity and other terms and how to investigate functional diversity. I think it's worth crediting the different approaches and the dissent. Some methods allow for an indirect approach to study functional diversity (those including taxa), others a direct approach (those including only functional groups). A review is a chance to point out methodological improvements over the last years and the review would gain relevance if some crosscompartment approaches are included here (e.g. specific comment #79).

Thank you for the critical comment. We have now added much more explanation in parts where the low specificity was pointed out. We have, e.g., explained biogeochemical mechanisms behind the presented hypotheses and added some methodological details, such as duration of experiments and size of micro- and mesocosms, to highlight the time and space scales. Suggested papers, and some others, are added.

We have written a new chapter 2, Definitions and review methods, where the key definitions used are explained.

We have included, in the chapter Knowledge gaps, a commentary on the potential usage of novel methods in climate change research in the Baltic Sea.

## Specific comments

- 1) p 1, ll. 3: add a definition of ecosystem functioning into your introduction or as a glossary, e.g. Ecosystem Functioning (Tilman 2001): The rate, level, or temporal dynamics of one or more ecosystem processes like primary production or nutrient gain or loss

A new full chapter 2, Definitions and review methods, has been written after the Introduction. The definitions are explained there.

- 2) p. 4, ll. 60-64: After Tilman (2001) the definition of ecosystem functioning (EF) is: The rate, level, or temporal dynamics of one or more ecosystem processes like primary production or nutrient gain or loss.

My question: How do you define EF in this review and which processes do you include in your review and which do you not include and why (e.g. are there other reviews out to refer to like "Wannicke, N., Frey, C., Law, C. S., & Voss, M. (2018). The response of the marine nitrogen cycle to ocean acidification. *Global Change Biology*, 24(11), 5031-5043.")? Pls add this information into your text.

Pls add a side note with definitions for the most prominent terms in your review, e.g.

- climate change
- ecosystem functioning
- trophic dynamics etc.

A new full chapter 2, Definitions and review methods, has been written after the Introduction. The approach and definitions are explained there.

- 3) p. 4, ll. 68: wording “...more light onto (not into) the complex...”

Corrected.

- 4) p. 4, ll 67-69: I understand that there is an upcoming review on the cc projections associated with this review. Still, for the "stand alone status" of this review it would be very helpful, to specify here in more detail the projected "abiotic" changes that possibly are most important for the biology (maybe less the atmospheric forcing behind them) including namely salinity, temperature, stratification and oxygen as well as OA, nitrate and phosphate levels, the latter from both rivers and the anoxic sediments. Important to include: How certain or uncertain can we be about them (e.g. in line 537-539)?

We have revised and expanded the Introduction thoroughly.

We have also included in Knowledge gaps more analysis of the consensus and dissensus, and uncertainties, and in Conclusions added more text on the possible attribution of the found responses to the global climate change and climatic variations in general.

Note also that there will be a separate review on Biogeochemistry of the Baltic Sea, so we only review biogeochemical processes if they are directly linked to the species and communities, or ecosystem functioning.

New text (in Introduction): “We primarily review studies that shed light to the effects of climate change on the Baltic Sea species, populations and communities, and the ecosystem function, by studying parameters which are assumed to change due to climate change, such as water temperature, salinity, oxygen, pH and nutrients. We also include studies that show past changes in communities, and those which study more indirect processes related to changes in biogeochemistry and food web, if these changes were attributed to parameters that have been shown to fluctuate with climatic variations. Most of these studies have in their Introduction referred to global climate change, and in their Discussion interpreted the results in the context of future climate change. Conclusions on the role of climate change on shaping the structure and functioning of the Baltic Sea ecosystem are then drawn from the existing field, experimental and modelling evidence.”

- 5) p. 4, ll. 70: add: „the“ before year

Corrected.

- 6) p. 4, ll. 71: add „field based“ before “responses”

Corrected.

- 7) p. 4, ll. 75: „food web dynamics“, what do you mean by this? Pls specify.

Edited text: “Third, the complex effects of climate change on the interactions between trophic groups, such as phytoplankton, cyanobacteria, zooplankton and fish, as well as algae or vascular plants and invertebrates inhabiting or grazing on them are analysed based on field and experimental studies where trophic interactions have been investigated.”

- 8) p. 4, ll. 75: add „both directly in the field as well as in experimental studies.” At the end of the sentence.

Added as suggested.

- 9) p. 4, ll. 76: What kind of modeling studies, pls specify.

Edited text: “Finally, a number of modelling studies, mostly based on coupled oceanographic-biogeochemical models, sometimes also including the main biotic components of the open sea

ecosystem, i.e., phytoplankton, cyanobacteria, zooplankton and planktivorous and piscivorous fish, as well as benthos, are reviewed.”

- 10) p. 4, ll. 85: Why don't you start with the autotrophic communities instead of the heterotrophic microbial community in Chapter 2?  
 Phototrophs form the base of the food web also for the heterotrophic microbial community that you seem to mainly refer to in Chapter 2.1.  
 Also, you should define, which organisms you mean with "microbial community", e.g. only heterotrophs?  
 What about marine viruses and fungi?

A valid comment. We now start with “Phytoplankton” and continue with “Cyanobacteria” (separated from the original chapter “Phytoplankton and Cyanobacteria”).

In the original chapter “Microbial communities” we referred to experimental studies investigating bacteria (not viruses or fungi), so we change the name of the Chapter to “Bacteria”, and move this chapter later in the paper (after the chapter Fish).

- 11) p. 4, ll. 87: add "(OA)" after ocean acidification

Added as suggested.

- 12) p. 4, ll. 90: How did community change? Be more specific.

Edited text: “OA alone had a limited impact, but when combined with increased temperature, certain bacterial phylotypes, such as betaproteobacteria, increased. The authors suggest that synergistic effects of increased temperature and acidification may selectively promote growth of specific bacterial populations.”

- 13) p. 4, ll. 91: Which OTUs? Be more specific.

Edited text: “In the southern Baltic Sea (Kiel Bight) the impact of OA was also limited to few operational taxonomic units (OTUs), such as *Bacteroidetes* ‘NS3a marine group’, as the bacterial community mainly responded to temperature and phytoplankton succession.”

- 14) p. 4, ll. 92: How did the microbial community respond? Be more specific.

New/edited text: “...the bacterial community mainly responded to temperature and phytoplankton succession. Depending on studied season and temperature treatment, Cyanobacteria, *Bacteroidetes*, Alphaproteobacteria and/or Gammaproteobacteria increased.”

- 15) p. 5, ll. 94: Give range also for CO<sub>2</sub> as for Sal and Temp.

Edited text: “In experiments using a natural summer microplanktonic community, where CO<sub>2</sub> was increased (from 380 to 960 μatm) and salinity decreased (from 6 to 3 psu),...”

- 16) p. 5, ll. 99 delete bracket before “Berner”

Unfortunately, the EndNote program forces this format. If the paper will be accepted, we will correct this at copy-editing stage.

- 17) p. 5, ll. 99 delete extra dot after et al.

There is no extra dot. It is a comma.

- 18) p. 5, ll. 100-101 Changed drastically to what? Be more specific. What does a "high temperature community" look like?

New/edited text: “At reduced salinity levels, certain Actinobacteria and Bacteroidetes OTUs increased, and the heterotrophic bacteria community resembled communities at high temperature, indicating synergistic effects of temperature and salinity.”

- 19) p. 5, ll. 102-103 Which increase? Unclear where you refer to here.

New/edited text: “Biotic interactions were more dominant than abiotic ones, however. The largest increase in heterotrophic bacterial biomass was detected when filamentous cyanobacteria started to decay, regardless of temperature or salinity. It was suggested that this indirect coupling between heterotrophic bacteria and filamentous cyanobacteria is more important for bacterial communities than the direct effects of temperature or salinity.”

- 20) p. 5, l. 104 (end of paragraph) What about the effect of other abiotic variables like stratification, oxygen, nitrate and phosphate on heterotrophic bacteria, viruses and fungi?  
Möller, L., Kreikemeyer, B., Gerdts, G., Jost, G., & Labrenz, M. (2021). Fish as a winter reservoir for *Vibrio* spp. in the southern Baltic Sea coast. *Journal of Marine Systems*, 221, 103574.  
Rojas-Jimenez, K., Rieck, A., Wurzbacher, C., Jürgens, K., Labrenz, M., & Grossart, H. P. (2019). A salinity threshold separating fungal communities in the Baltic Sea. *Frontiers in microbiology*, 10, 680.

New/edited text: “This highlights the importance of considering both abiotic and nutrient effects and the more indirect food web effects, i.e. predation, on microbial communities.”

The papers of Möller et al. (2021) and Rojas-Jimenez et al. (2019) are interesting but are not specifically referring to climate change, so we chose not to include them.

- 21) p. 5, ll. 106 Cyanobacteria is a wide field, namely in the BS. The ecology can be very different. Some are palatable (unicells also called picocyanobacteria seem to be palatable for mesozooplankton), some are not (the large, filamentous ones are hardly grazed directly, right?). So pls refine what you mean by cyanobacteria, and why they may (or may not) be problematic.

We are mainly reviewing papers that study filamentous, bloom-forming cyanobacteria, and make this clear in relevant places, especially when considering zooplankton grazing.

To make the text better structured, we have separated text on cyanobacteria from the chapters “Phytoplankton and cyanobacteria”, and “Climate change effects on primary producers”, to a new chapter “Cyanobacteria”.

- 22) p. 5, ll. 109 Be more specific: not few but last 30? years?  
23) p. 5, l. 110 Although with a clear gap between spring and summer blooms, right?  
24) p. 5, ll. 112-116 Is that true for all basins of the BS?

Excellent points. As an answer to comments 22-24 we have thoroughly edited the text in this point.

New/edited text: “The growing season of phytoplankton has significantly prolonged with warming temperatures during the past few decades. A satellite-based study suggested that the period with chlorophyll-a of at least 3 mg m<sup>-3</sup> doubled, from 110 days in 1998 to 220 days in 2013 (Kahru et al., 2016). Another study using phytoplankton sampling data from the shallow Bay of Mecklenburg, western Baltic Sea, confirmed that the phytoplankton growing season, which in 1988-1992 on average endured from March to August, now (2014-2017) extends from February to December (Wasmund et al., 2019), albeit with a longer gap between the spring and late summer peaks. This prolongation was tentatively explained by increased sunshine in spring and higher temperature in the autumn, inducing changes in various factors, such as species composition and settling rates of phytoplankton, remineralization of organic matter by bacteria, and grazing rates by zooplankton (Wasmund et al., 2019). Although studies from all Baltic Sea basins do not exist, it is probable that similar prolongation, caused by changes in radiation and temperature, have also taken place elsewhere.”

- 25) p. 5, l. 115 add summarized by: Spilling, K., Olli, K., Lehtoranta, J., Kremp, A., Tedesco, L., Tamelander, T., ... & Tamminen, T. (2018). Shifting diatom—dinoflagellate dominance during spring bloom in the Baltic Sea and its potential effects on biogeochemical cycling. *Frontiers in Marine Science*, 5, 327.

**Reference added.**

Consider adding: Paul, A. J., Sommer, U., Paul, C., & Riebesell, U. (2018). Baltic Sea diazotrophic cyanobacterium is negatively affected by acidification and warming. *Marine Ecology Progress Series*, 598, 49-60.

**Reference added, and a notion made on the discrepancy between modelling and mesocosm studies regarding potential increase of Cyanobacteria.**

**New text:** “It is also notable that, in mesocosm studies, an increase of pCO<sub>2</sub> (from 360 to 2030 µatm) coupled with an increase in water temperature (from 16.6 to 22.4 °C) had a *negative* impact on the biomass of the diazotrophic cyanobacteria *Nodularia spumigena* (in 1400-L mesocosms, 28 days) (Paul et al., 2018). This result contradicts the modelling studies that suggest that the increased stratification, together with potentially increasing remineralization of organic matter and release of phosphorus from the anoxic sediments, will increase cyanobacteria blooms (Meier et al., 2011a; Andersson et al., 2015; Neumann et al., 2012; Chust et al., 2014).”

- 26) p. 5, l. 117 Careful: There are hardly Cyanobacteria in the western BS. Pls add the basins that you refer to as this is unclear from the title of your review, which includes the whole BS, not just the Baltic Proper and adjacent gulfs.

**A valid comment. We have added notions on the study areas in several places in the ms.**

**New/edited text (in the chapter Phytoplankton):** “In the northern Baltic Proper, Åland Sea and the Gulf of Finland, the biomasses of Chrysophyceae, Prymnesiophyceae and Cyanophyceae have increased (Suikkanen et al., 2013), and the phytoplankton biomass maximum, which in the 1980’s was in spring and mainly consisted of diatoms, is now in July-August and is dominated by filamentous cyanobacteria.”

- 27) p. 5, l. 118 add: "comprising of diatoms" after “spring bloom”  
 28) p. 5, l. 118 add: "comprising of mainly unpalatable cyanobacteria" after “August”

**Edited text:** “and the phytoplankton biomass maximum, which in the 1980’s was in spring and mainly consisted of diatoms, is now in July-August and is dominated by filamentous cyanobacteria.”

- 29) p. 5, ll. 117-121 Here the text is unclear where you refer to the spring and summer blooms, respectively. The top down pressure probably refers to the spring bloom, as cyanobacteria are hardly grazed directly, right? This does not become clear in the text at the moment. Pls rephrase for clarity, e.g., by adding "on the diatom blooms in spring"

**Edited text:** “This shift has been explained by a complex interaction between warming, eutrophication and increased top-down pressure on species of the spring bloom, as well as changes in DIN:DIP ratio in summer (Suikkanen et al., 2013).”

- 30) p. 5, ll. 120-121 You leave out the most interesting info here: change from which phytoplankton group to which other phytoplankton group?

**New text (in the chapter Cyanobacteria):** “Also, in the Gulf of Bothnia, changes in phytoplankton and cyanobacteria communities have been observed. In the Bothnian sea, concentration of chlorophyll *a* has increased in summer, along with the increase of cyanobacteria and the mixotrophic ciliate *Mesodinium rubrum* (Kuosa et al., 2017).”



- 31) p. 5, ll. 122-125 Consider moving into section 3.1. Climate change and primary production in the pelagial

Thank you for the suggestion for restructuring the text. Sentence moved as suggested. We have however collated most text concerning cyanobacteria under a new chapter “Cyanobacteria”, to avoid overlap between chapters “Phytoplankton and cyanobacteria” and “Climate change and primary production”.

- 32) p. 5, ll. 122-123 Again: be more specific, which changes in pelagic PP do you mean?

New/edited text (in chapter “Projections of primary producers”, formerly called Climate change and primary productivity): “Experimental (mesocosm) evidence also supports findings that climate change induced warming up of water and changes in light conditions will drive changes in the pelagic primary producers, by accelerating spring bloom, inducing a decline in peak biomass and favouring small size cells, either directly or via increased grazing by copepods (Sommer et al., 2012).”

- 33) Nummer: 20 l. 124 Be more specific: Which ecosystem wide consequences do you mean?

New/edited text (in chapter “Projections of primary producers”): “A thorough review illustrating benthic-pelagic coupling shows ecosystem-wide consequences of altered pelagic primary production, e.g. via increasing sedimentation of organic matter and consequent remineralization, inducing hypoxic conditions both in the deep basins and in the shallower archipelago areas (Griffiths et al., 2017), probably also inducing internal loading of phosphorus from sediments (Puttonen et al., 2014; Stigebrandt et al., 2014)...”

- 34) p. 5, l. 125 What exactly do you mean with "food web dynamics"?

Edited text: “...and impacting abundances and interactions between main trophic levels, e.g. phytoplankton, detritus and zoobenthos as well as detritivores, benthivores phytoplanktivores, zooplanktivores and piscivores (Kortsch et al., 2021).”

- 35) p. 5, l. 126 And with "climate" you mean what again?

We use it in the same meaning as the papers cited in our ms. We have added a separate Chapter 2 – “Definitions and review methods”, where we explain our own definitions.

Edited text: “There is, however, a discrepancy on the relative effects of eutrophication and climate change in explaining past variations in phytoplankton communities and biomass.”

- 36) p. 5, l. 125 add BS literature: Kiljunen, M., Peltonen, H., Lehtiniemi, M., Uusitalo, L., Sinisalo, T., Norkko, J., ... & Karjalainen, J. (2020). Benthic-pelagic coupling and trophic relationships in northern Baltic Sea food webs. *Limnology and Oceanography*, 65(8), 1706-1722

Reference added.

- 37) p. 5, l. 124 Again you talk about changes without specifying which phytoplankton groups were replaced by which other groups. Pls specify.

On line 124 we did not refer to specific phytoplankton groups. Edited as explained in replies to comment numbers 33 and 34.

- 38) p. 5, l. 127 switch community and biomass as so far you have mainly talked about community

Edited as suggested.

- 39) p. 5, l. 128 Give an example for dominant species, pls.

New text: “For instance, spring phytoplankton biomass increased in the Baltic Proper and decreased in the Belt Sea area, both areas showing oscillations between communities dominated by diatoms or dinoflagellates (Wasmund et al., 2011).”

- 40) p. 5, l. 128 Dominating in density (aka abundance) or biomass?

We refer to biomass. See answer to Comment no. 39.

- 41) p. 5, l. 129 Add something like „...leading to a switch from group x to group y.” after effect. Pls be more specific.

We agree that this paragraph was not specific enough, and we have now completely rewritten this chapter.

New/edited text: “A study comparing historic phytoplankton communities from 1903-1911 with the present ones (1993-2005) in the northern Baltic Proper and the Gulf of Finland observed an undefined “period effect”, characterized by a decline of diatoms and increase of dinoflagellates, that was not well explained by the available environmental variables (temperature, salinity and climatological data). Although data on biogeochemical parameters was not available for the period 1903-1911, the authors interpreted the observed community change as evidence of the direct and/or indirect influence of eutrophication (Hällfors et al., 2013).

A fifteen-year study (2000-2014) using FerryBox observations, covering the area between Helsinki (Gulf of Finland) and Travemünde (Mecklenburg Bight), confirmed that spring bloom intensity was mainly determined by winter nutrient concentration, while bloom timing and duration co-varied with meteorological conditions. The authors conclude that the bloom magnitude has been affected by the reduction of nutrient loading from land, while bloom phenology can also be modified by global climate change affecting seasonal oceanographic and biogeochemical processes (Groetsch et al., 2016). It has also been noted that the trends in certain groups, like cryptophytes, may be affected by anomalies in the Baltic Sea Index, a regional climate index similar to NAO, although a mechanistic explanation for the relationship could not be found (Griffiths et al., 2020).”

- 42) p. 6, l. 131 Pls define BSI

New text: “It has also been noted that the trends in certain groups, like cryptophytes, may be affected by anomalies in the Baltic Sea Index, a regional climate index similar to NAO, although a mechanistic explanation for the relationship could not be found (Griffiths et al., 2020).”

- 43) p. 6, l. 132 Pls, explain which change you mean, e.g. be more specific. Otherwise the reader has no clue of what quality the changes are that you review about.

- 44) p. 6, l. 132 Density or biomass (e.g. cell-carbon) wise community changes?

Edited text: “Other studies did not find any explanation for the observed changes in the biovolumes of different taxa, e.g. decrease of diatoms and increase of certain dinoflagellate taxa, and concluded that phytoplankton community in the Baltic Sea is not in a steady state (Olli et al., 2011),...”

- 45) p. 6, l. 138 Do you mean phytoplankton or cyanobacteria or both?

The cited study does not make this distinction.

Edited text: “...and together with increased internal loading of nutrients (Stigebrandt et al., 2014), several modelling studies project an increase in total phytoplankton concentration (in mgChl m<sup>-3</sup>) until the end of the century (Meier et al., 2012a; Meier et al., 2012b; Skogen et al., 2014; Ryabchenko et al., 2016).”

- 46) p. 6, ll. 140-141 How does that mechanism work? Pls explain in more detail. Really, evidence? Or rather indication? What kind of evidence do you refer to? Also, do you mean predicted or projected climate change?

We have now edited the text thoroughly and explained the process affecting phytoplankton more carefully.

We move the rest of the description of the Kattegat regime shift to the Chapter Climate change and regime shifts.

New/edited text (in this chapter): “This has also been seen in Kattegatt, where reduction of nutrient loading led in mid 1990s to a shift from a highly eutrophic state characterized by small phytoplankton species and low water transparency to increasing share of diatoms, decreasing overall phytoplankton biomass and increase of water transparency (Lindegren et al., 2012). The improving oxygen conditions and increase of water temperature, induced by cyclic climatic variations (positive NAO) and gradual warming of climate, respectively, mainly affected the benthic ecosystem (Lindegren et al., 2012).”

New text (in Climate change and regime shifts): “Also in Kattegat, the western Baltic Sea, where the ecosystem is more oceanic than in the Baltic proper and the northern Baltic Sea, a drastic regime shift was detected in mid 1990s. First, a drastic reduction of nutrient loading, led into a shift from a highly eutrophic, pelagic ecosystem state to an ecosystem characterized by decreasing overall phytoplankton and meso- and microzooplankton biomass, dominance by small sized fish in the pelagial, an increase of macroalgae and filter-feeding molluscs in the hard bottoms and other benthic animals in the soft sediments (Lindegren et al., 2012). Second, climate variability, i.e., positive NAO and Baltic Sea index (a regional climate index), has increased inflow of well oxygenized water from the North Sea into the area, improving conditions for zoobenthos, including, e.g., populations of the commercially important Norway lobster. Further, increase of sea surface temperatures, possibly induced by global climate change, probably has contributed to the improved flatfish growth and survival in the shallow nursery areas (Lindegren et al., 2012). Decreasing fishing may also have been contributed to the increase of gadoids and flatfish, but its relative importance is difficult to distinguish from other co-occurring effects. However, it is obvious that regime shifts are often a result of several environmental, climatic and anthropogenic effects acting synergistically on the entire ecosystem.”

- 47) p. 6, ll. 142-145 Consider moving into section 3.1. Climate change and primary production in the pelagial

Thank you for the comment. We notice now that there is much overlap between chapters “Phytoplankton and cyanobacteria” and “Climate change and primary production in the pelagial”. We decided to move part of the contents of the latter chapter to separate chapters “Phytoplankton” and “Cyanobacteria”. These two chapters are now rewritten, to give a more coherent picture concerning pelagic primary producers.

- 48) p. 6, ll. 143-145 Pls revise sentence for correct grammar.

Grammar corrected.

- 49) p. 6, ll. 143-145 Add detailed info, e.g.: “..increases by xy% compared to the control of ambient conditions.”

Edited text: “E.g. the biomass of southern Baltic autumn phytoplankton (kept in 1400-L indoor mesocosms for 21 days) increased when pCO<sub>2</sub> was increased from 439 ppm to 1040 ppm, also under warm conditions (Sommer et al., 2015).”

- 50) p. 6, l. 143 Add „the“ before „water“

Added as suggested.

- 51) p. 6, l. 148 Wording: „decrease in” or “release in grazing pressure from”, your choice.

Wording changed to: “caused by an associated decrease of grazing by copepod nauplii.”

52) p. 6, ll. 150-154 Long sentence. Pls break up in shorter ones for better readability.

Sentence split in two.

53) p. 6, l. 150 Have you defined at the beginning of your review, what you mean anytime you say CC? If not, pls add. If CC includes different variables in different parts of the text, I think you need to specify, which variables you refer to in each case.

A valid comment. We have now added a chapter 2 – Definitions and review methods after the Introduction. We explain all main definitions there.

54) p. 6, l. 151 add Jerney, J., S. Suikkanen, E. Lindehoff and A. Kremp (2019). Future temperature and salinity do not exert selection pressure on cyst germination of a toxic phytoplankton species. *Ecol. Evol.* 9: 4443-4451, doi: 10.1002/ece3.5009

Reference added.

55) p. 6, ll. 150-154 convoluted sentence, pls revise for clarity.

Sentence split in two (cf. comment no. 52)

56) p. 6, ll. 154-155 I don't think cyanobacteria per se are a problem. Unicells should not be a problem, right? Yet they are cyanobacteria. The problem are large, unpalatable Cyanobacteria, which should be specified here and elsewhere.

A valid point. New/edited text: “As toxins of both dinoflagellates (Sopanen et al., 2011) and cyanobacteria (Karjalainen et al., 2006; Karjalainen et al., 2007; Engström-Öst et al., 2017) can accumulate in Baltic Sea zooplankton and induce lower grazing rates and higher mortality, these studies suggest that toxic dinoflagellates and filamentous unpalatable cyanobacteria may get a competitive advantage against diatoms in a future Baltic Sea.”

57) p. 6, l. 155 Why another? Which is the first competitive advantage to begin with that you seem to refer to?

Phrase “yet another” is deleted.

58) p. 6, ll. 154-155 This is contradictory to what is stated in ll. 603-605.

A valid point. Therefore, we delete “yet another” (see response to comment no. 57). The sentence on lines 603-605 refers to studies that note that reducing nutrient loading according to BSAP would decrease the intensity of cyanobacteria blooms in the future. These are two counteracting effects, but it is obvious that modelling studies cannot presently take into account complex biological interactions between species. This is now highlighted as one of the knowledge gaps.

59) p. 6, l. 157 Consider the field study: Eglite, E., Wodarg, D., Dutz, J., Wasmund, N., Nausch, G., Liskow, I., et al. (2018). Strategies of amino acid supply in mesozooplankton during cyanobacteria blooms: A stable nitrogen isotope approach. *Ecosphere*, 9, e02135. <https://doi.org/10.1002/ecs2.2135>

Thank you for the suggestion, reference added.

New text: “Sufficient supply of essential compounds such as amino acids (AA) produced by phytoplankton and cyanobacteria is essential for the growth and productivity of zooplankton grazers. It has been suggested that this supply may change if the sea surface temperature increases. A field study performed in the Baltic Proper explored the natural abundances of AA in particulate organic matter and mesozooplankton (Eglite et al., 2018). The results show that, during a warm summer, thermophilic rotifers and cladocerans (e.g. *Bosmina* spp.) acquired ample AA through filter feeding on the abundant diazotrophic cyanobacteria, whereas the temperate copepods (e.g. copepods *Temora longicornis* and *Pseudocalanus* spp.) avoided the warm surface

layer and acquired AA mainly through sinking organic matter and/or via grazing on chemoautotroph based microbial food web in the suboxic zone. This may imply that thermophilic zooplankton species, such as rotifers and certain cladocerans gain more AA than copepods in a future warmer and more stratified Baltic Sea.”

60) p. 6, ll. 159-161 Like whom? Pls add species.

Species added. New/edited text: “Several studies have confirmed that during the 1980s and 1990s marine copepod species (e.g., *Pseudocalanus* spp. and *Temora longicornis*) declined, while euryhaline and limnetic, smaller-sized copepod species (*Acartia* spp. And *Eurytemora* spp.) increased in abundance (Hänninen et al., 2015; Suikkanen et al., 2013).”

61) p. 6, l. 160 add "copepod" after small-sized

Added as suggested.

62) p. 6, l. 160: delete the comma after „small sized“

Deleted as suggested.

63) p. 6, l. 161 Give examples of marine taxa.

64) p. 6, l. 162 Give examples for brackish-water taxa.

Examples given. See reply to comment no. 60.

65) p. 6, l. 164 add the following study to explain at least one underlying mechanism: Dutz, J., & Christensen, A. M. (2018). Broad plasticity in the salinity tolerance of a marine copepod species, *Acartia longiremis*, in the Baltic Sea. *Journal of Plankton Research*, 40(3), 342–355.  
<https://doi.org/10.1093/plankt/fby013>

Thank you for the suggestion. Reference added.

New text: “Also, it has been experimentally shown that close to the physiological tolerance limit for salinity, respiration of copepods (*Acartia longiremis*) increases and feeding rate decreases (in 610 ml bottles, 24 h experiments), indicating a disruption of the energetic balance under low salinity (Dutz and Christensen, 2018).”

66) p. 7, l. 167: Small scale impacts like what? Pls be more specific.

Edited text: “Environmental impacts on the physiology of the more sensitive species may however affect their reproductive success, and thus influence both populations and communities (Möller et al., 2015).”

67) p. 7, l. 174: In which way are cladocerans and rotifers different functional groups e.g. after the definition of Tilman (2001)? They are different taxonomic groups but as stated it is not clear why they would represent a functional group. E.g. which specific function do they represent? Also: “Shift” from whom? Herbivory, omnivory or carnivory are functional groups. Do you mean eventually a shift from omnivorous copepods to herbivorous cladocerans and rotifers? I’m confused, pls clarify. Again, a definition how you define “functional group”, “functional diversity” etc is urgently needed in this review. Again, I suggest definitions of terms as given in the glossary of “Tilman, D. (2001). Functional diversity. *Encyclopedia of biodiversity*, 3(1), 109-120.”

A valid comment. We here refer to filter-feeding vs. raptorially feeding species, not herbivorous or carnivorous. We made an attempt to clarify the processes better.

New/edited text: “Changes in zooplankton functional groups, such as a shift from raptorially and suspension-feeding copepods and cladocerans to a dominance by filter-feeding rotifers and cladocerans, have been shown as a result of warming (Suikkanen et al., 2013; Jansson et al., 2020). Also, a switch from predominantly herbivorous feeding to carnivory (feeding on ciliates)

has been observed in a field study in the southern and central Baltic Sea, during cyanobacterial blooms (Loick-Wilde et al., 2019), probably supported by decomposing of the otherwise unpalatable filamentous cyanobacteria, and an associated increase of the heterotrophic pathways of energy (Hogfors et al., 2014). It is possible that the functions of zooplankton community will change as climate-induced warming and reduced salinity continues.”

68) p. 11, l. 337 What about food quality and transfer efficiency of mass and energy?

Food quality added. We consider transfer efficiency in the chapter “Nutrient cycling, benthic pelagic coupling and trophic efficiency”.

Edited text: “Future climatic variations may affect fish in the Baltic Sea through their effects on water temperature, salinity, oxygen and pH, as well as nutrients, which indirectly affect availability and quality of food for fish.”

69) p. 11, l. 338: After „fish“ add: Limburg, K. E., & Casini, M. (2019). Otolith chemistry indicates recent worsened Baltic cod condition is linked to hypoxia exposure. *Biology letters*, 15(12), 20190352. Möllmann C, Cormon X, Funk S, Otto SA, Schmidt J, Schwermer H, Sguotti C, Voss R, Quaas M (2021): Tipping point realized in cod fishery, *Nature Scientific Reports*, DOI: <https://doi.org/10.1038/s41598-021-93843-z>

Thank you for the suggestion. References added.

70) p. 13, l. 390: Consider adding the following study to determine trophic efficiency in field samples, because it includes examples also from the Oeland upwelling and larger Baltic Proper in the Fig. 6 : Weber, S. C., Loick-Wilde, N., Montoya, J. P., Bach, M., Doan-Nhu, H., Subramaniam, A., ... & Voss, M. (2021). Environmental regulation of the nitrogen supply, mean trophic position, and trophic enrichment of mesozooplankton in the Mekong River plume and southern South China Sea. *Journal of Geophysical Research: Oceans*, 126(8), e2020JC017110. namely Chapter:”4.4. Ecosystem-Specific Trophic Enrichment in Mesozooplankton”

Thank you for the suggestion. We however want to concentrate citing papers that have a focus on the Baltic Sea, and therefore prefer not to include this study.

71) p. 13, l. 390: trophic efficiency: Pls define in the text or glossary

Defined in chapter 2 – Definitions and review methods.

72) p. 13, l. 390 At the moment it does not become clear why some processes are reviewed and others are missing. E.g. what do we know about key processes like nitrification, denitrification, N<sub>2</sub> fixation in a future Baltic Sea?

Suggested References from whichs´results at least some clues may be deduced:

Bartl, I., Hellemann, D., Rabouille, C., Schulz, K., Tallberg, P., Hietanen, S., & Voss, M. (2019).

Particulate organic matter controls benthic microbial N retention and N removal in contrasting estuaries of the Baltic Sea. *Biogeosciences*, 16(18), 3543-3564.

Allin, A., Schernewski, G., Friedland, R., Neumann, T., & Radtke, H. (2017). Climate change effects on denitrification and associated avoidance costs in three Baltic river basin-coastal sea systems. *Journal of Coastal Conservation*, 21(4), 561-569.

Asmala, E., Carstensen, J., Conley, D. J., Slomp, C. P., Stadmark, J., & Voss, M. (2017). Efficiency of the coastal filter: Nitrogen and phosphorus removal in the Baltic Sea. *Limnology and Oceanography*, 62(S1), S222-S238.

Hellemann, D., Tallberg, P., Bartl, I., Voss, M., & Hietanen, S. (2017). Denitrification in an oligotrophic estuary: a delayed sink for riverine nitrate. *Marine Ecology Progress Series*, 583, 63-80.

Olofsson, M., Klawonn, I., & Karlson, B. (2021). Nitrogen fixation estimates for the Baltic Sea indicate high rates for the previously overlooked Bothnian Sea. *Ambio*, 50(1), 203-214.

Loick-Wilde, N., Weber, S. C., Eglite, E., Liskow, I., Schulz-Bull, D., Wasmund, N., ... & Montoya, J. P. (2018). De novo amino acid synthesis and turnover during N<sub>2</sub> fixation. *Limnology and Oceanography*, 63(3), 1076-1092.

Thank you for the suggestions. We have not reviewed biogeochemical processes thoroughly, because in the ESD BEAR Special Issue, there is a separate manuscript, [Kulinski et al., on biogeochemistry of the Baltic Sea https://esd.copernicus.org/preprints/esd-2021-33/esd-2021-33.pdf](https://esd.copernicus.org/preprints/esd-2021-33/esd-2021-33.pdf) : Almost all the suggested papers are reviewed in that paper. In our review we have highlighted mainly papers that connect biogeochemical processes directly to biota. We therefore choose not to cite the suggested papers in our review.

73) p. 13, l. 412-415: This is interesting! Pls explain the mechanism behind this at least briefly.

New/edited text: “It has also been suggested that climate change may decrease fish productivity, especially in the northernmost Baltic Sea, because, when the system shifts towards heterotrophy, the food web efficiency declines, due to competition for nutrients between bacteria and phytoplankton, and the phytoplankton production decreases. This creates less food for zooplankton and planktivorous fish, and also decreases sedimentation of organic matter, reducing benthic production and diminishing food availability for benthic-eating fish. Eventually the fish production may decrease (Berglund et al., 2007; Wikner and Andersson, 2012).”

74) [No comment under this number.]

75) p. 13, l. 413: Add „switches“ after „system“

Corrected, albeit with the word “shifts”.

76) p. 13, ll. 415-418: I don't understand this. If the food web bases on heterotrophy rather than photoautotrophy, doesn't that imply that less mass and energy is transferred to fish since the lower food chain is elongated (based on heterotrophs rather than autotrophs) leading to a higher trophic position in mesozooplankton (e.g. carnivorous zoops instead of herbivorous zoops, e.g. as documented Loick-Wilde et al. 2019)? Then e.g. eastern Baltic cod would also have a higher TP in such an, at times, heterotrophy based system, compared to a lower TP in western Baltic cod due to a phototrophy based system, right? Pls clarify.

We agree that the wording was not clear enough. We have now revised the text to better explain the rationale behind the idea of microbial loop sustaining high zooplankton and fish production, and the associated uncertainties.

New/edited text: “Certain mesocosm studies simulating effects of climate change have however found that the production and biomass of copepods can remain high, even when they feed upon the longer bacteria-flagellate-ciliate food chain, because the positive effects of increasing temperature on copepod production override the negative effects of decreasing food web efficiency (Lefebure et al., 2013). Furthermore, many Baltic Sea copepods are omnivorous and can opportunistically switch between suspension feeding on flagellates and raptorial feeding on ciliates (Kiorboe et al., 1996). This creates an ‘intraguild’ relationship between the three trophic levels, flagellates, ciliates and copepods (Gismervik and Andersen, 1997), which stabilizes the system and can sustain copepod production even under lower phytoplankton production. If copepod production remains high, also fish production may be supported also when the system shifts to heterotrophic production (Lefebure et al., 2013). On the other hand, a study performed in a large biotest area artificially heated by the cooling waters of the Forsmark Nuclear power plant, southern Bothnian Sea, found that warming of water may lead to increased species turnover, and in decreased compositional stability of diatom, macrophyte and invertebrate communities (Hillebrand et al., 2010). As it is challenging to incorporate such complex interactions in 3D ecosystem models, the consequences of climate change on trophic efficiency and future fish production under different scenarios and in different sea areas remain unsecure.”

- 77) p. 15, ll. 461-467: I disagree with “warming induces a switch from a bottom-up controlled to a mainly top-down controlled system, which may result in increased zooplankton abundance and reduced phytoplankton biomass under warm temperature”. What consequences do you think it has for higher trophic levels like fish or sea birds if mesozooplankton switches from herbivory to carnivory due to increasing densities of unpalatable cyanobacteria in a future Baltic Sea? According to a simple biogeochemical model (see Figure below from T&T 2011), the decrease in mass and energy that is available for TPs above the mesozooplankton compartment should be massive, shouldn't it? Pls discuss in a larger context.

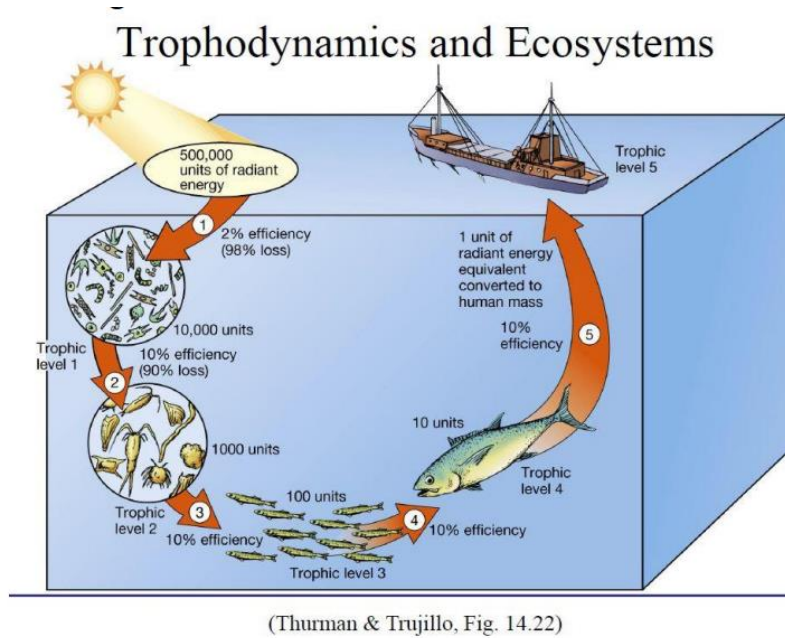


Figure 14.22 shows the passage of energy between trophic levels through an entire ecosystem, from the solar energy assimilated by the autotrophic plankton through all trophic levels to piscivorous humans. From the assimilated chemical energy, a large fraction is converted by respiration into kinetic energy for sustaining life or is lost as heat, and what remains is available for growth and reproduction. Thus, only about 10% of the ingested by herbivores is available for the next trophic level. Since energy is lost at each trophic level, it takes thousands of smaller marine organisms to produce a single fish that can be so easily consumed during a meal! Source: Trujillo, A. P., & Thurman, H. V. (2011). *Essentials of oceanography* (10th edition). Pearson Education. pp. 551.

We agree that the wording was not clear, and we have now rewritten this part to better explain the suggestions of the cited studies. In this chapter we avoid judging the validity of published studies, but try to review their rationale as best as we can.

New/edited text: “In areas where climate change increases the supply of allochthonous DOM into the system, and warms up the sea surface temperature, strengthening the stratification and reducing the availability of nutrients from deeper waters, phytoplankton production may decline and the trophic pathways from bacteria and flagellates through ciliates to copepods may strengthen (Aberle et al., 2015). It has also been suggested, from experimental (mesocosm) evidence, that warming speeds up the growth of copepods but leaves phytoplankton unaffected, which shortens the time lag between phyto- and zooplankton. This may lead to a larger and earlier zooplankton peak and increase the possibility of zooplankton controlling phytoplankton, which may lead to a reduced phytoplankton biomass under warm temperature (Paul et al., 2016). Such results highlight the importance of considering food web effects (both bottom-up and top-down) on the pelagic ecosystem under climate change.”

- 78) p. 15, ll. 461-467: Add the following field based study about the environmental regulations of a switch from herbivory to carnivory in mesozooplankton in the Baltic Sea in summer in this paragraph: Loick-Wilde, N., Fernandez-Urruzola, I., Eglite, E., Liskow, I., Nausch, M., Schulz-Bull, D., et al. (2019). Stratification, nitrogen fixation, and cyanobacterial bloom stage regulate the



planktonic food web structure. *Global Change Biology*, 25(3), 794–810. <https://doi.org/10.1111/gcb.14546>.

Reference added. New/edited text: “Changes in zooplankton functional groups, such as a shift from raptorially and suspension-feeding copepods and cladocerans to a dominance by filter-feeding rotifers and cladocerans, have been shown as a result of warming (Suikkanen et al., 2013; Jansson et al., 2020). Also, a switch from predominantly herbivorous feeding to carnivory (feeding on ciliates) has been observed in a field study in the southern and central Baltic Sea, during cyanobacterial blooms (Loick-Wilde et al., 2019), probably supported by decomposing of the otherwise unpalatable filamentous cyanobacteria, and an associated increase of the heterotrophic pathways of energy (Hogfors et al., 2014).”

- 79) p. 17, l. 535: There is also a significant knowledge gap about the chances of new methodological approaches. So how about including methodological improvements that allow for a significant reduction in trait complexity while considering intraspecific variations in biological samples and specifically allow for the calibration and validation of current biogeochemical models?

Using compound-specific isotope analyses (CSIA) of amino acid nitrogen, it is now possible to measure a continuous trophic position in any biological compartment (as opposed to discrete trophic levels) based on a single field sample, which integrates the assimilation of mass from all the trophic pathways leading to a top predator from different field locations. With this information, we can take the next step of relating the effective TPs e.g. of zooplankton to the environmental conditions measured in situ (Loick-Wilde et al., 2019), providing much needed insights into the mechanisms driving shifts in TP.

The strength of CSIA lies in providing information on both TP and N sources from a single organism/sample, which is achieved with a simple comparison of the  $\delta^{15}\text{N}$  values of glutamic acid (Glu) and phenylalanine (Phe) amino acids (McClelland & Montoya, 2002; Mompean et al., 2016). While Glu is enriched in  $^{15}\text{N}$  by  $\sim 8.0\%$  per trophic transfer (Chikaraishi et al., 2009), the  $\delta^{15}\text{N}$  of Phe remains nearly unchanged when the amino acid (AA) is transferred through the food web and thus reflects the isotopic composition of the primary producers (Nsource measure, Chikaraishi et al., 2010). This approach largely eliminates potential sources of error in TP estimates associated with temporal and physiological decoupling between a consumer and its diet, and has been refined and confirmed in numerous field- and lab-based trophic studies over the last decade (reviewed by Glibert et al., 2019 and Ohkouchi et al., 2017).

The CSIA based N source identification and mean trophic position from cross-compartment analyses can directly be used to calibrate and validate current biogeochemical models and allow for an end-to-end quantification e.g. of N inputs from  $\text{N}_2$  fixation into apex predators like cod or sea birds.

A good point and we appreciate the suggestion. We have added a notion on novel methods in Knowledge gaps.

New text: “Also, more extensive use of biochemical and genetic methods, such as biomarkers (Turja et al., 2014; Turja et al., 2015; Villnäs et al., 2019), stable isotopes (Voss et al., 2000; Gorokhova et al., 2005; Morkune et al., 2016; Lienart et al., 2021), compound-specific isotope analyses (Ek et al., 2018; Weber et al., 2021) or metabarcoding (Leray and Knowlton, 2015; Bucklin et al., 2016; Klunder et al., 2021) could yield novel information on community structure, stress levels experienced by organisms, and of the trophic position of various taxa, under environmental change. Such information would allow validation of current biogeochemical models under different environmental scenarios, including climate change.”

- 80) p. 17, l. 535: Why do you refer only to salinity and stratification, what about the other abiotic variables like temperature, oxygen, OA, nitrate or phosphate inputs (either through rivers or from the anoxic sediments)?

A valid comment. New/edited text: “Projections of sea surface temperature and ice conditions are held relatively reliable, but, despite more than two decades of 3D modelling, there are still large

uncertainties in projecting certain basic physical parameters like salinity level as well as stratification under different climate forcings. Consequently, it is also difficult to project all parameters affected by stratification, especially oxygen levels and release of nutrients from the sediments, during different periods, at different depths and in different sea areas. Also, long term variation in external loading of nutrients from rivers depends, in addition to the magnitude of anthropogenic loading, also biogeochemical processes in the soil and in lakes and rivers. All these uncertainties weaken our ability to project marine biological processes, from pelagic primary and secondary productivity and benthic-pelagic coupling to zooplankton and fish populations, and to geographic shifts in macroalgal communities and invertebrates inhabiting the photic zone.”

- 81) p. 19, l. 604: The following review is less certain about a future decrease in cyanobacteria, pls discuss more controversial: Munkes, B., Löptien, U., & Dietze, H. (2021). Cyanobacteria blooms in the Baltic Sea: a review of models and facts. *Biogeosciences*, 18(7), 2347-2378.

What about the internal P storage in the sediments that gets released under anoxic conditions? Stigebrandt, A., Rahm, L., Viktorsson, L., Ödalen, M., Hall, P. O., & Liljebladh, B. (2014). A new phosphorus paradigm for the Baltic proper. *Ambio*, 43(5), 634-643.

Thank you for the comment. The reference is added to the chapter on Cyanobacteria. Citations on Stigebrandt et al. (2014) are inserted in several places where internal loading is dealt with.

New/edited text: Intensified blooms of cyanobacteria are expected especially if hypoxia will prevail and internal loading will increase supply of phosphorus from anoxic sediments into the surface layer, decreasing the N:P ratio (Meier et al., 2011b; Funkey et al., 2014). However, the magnitude of changes and their consequences for biogeochemical processes, e.g. for nitrogen fixation, differ greatly between models (Munkes et al., 2021).”

- 82) p. 19, l. 608: What about empirical field observations/ research? Pls add.

Thank You for the suggestion. Edited text: “These studies further highlight the importance of studying the Baltic Sea as a socio-ecological system, responding to both environmental and societal changes (Bauer et al., 2018; Bauer et al., 2019; Hyytiäinen et al., 2019), and it is important to continue efforts combining empirical field studies, experimental studies, modelling and dialogue with human society in order to attune to the changes ultimately driven by the Ocean itself (Stenseth et al., 2020).”

- 83) p. 20, l. 623: add any missing field and lab studies as pointed out in the text here, too, when applicable.

The Table 1 is updated to include all papers suggested, and some more.

## Technical corrections

Just few, part of the specific comments!

A few typos corrected and some grammatical improvements made.